

169.0018

**TITLE OF THE INVENTION**

**INTRON SEQUENCE ANALYSIS METHOD  
FOR DETECTION OF ADJACENT AND REMOTE LOCUS ALLELES  
AS HAPLOTYPES**

169.0018

CROSS REFERENCE TO RELATED APPLICATION

This application is a Continuation In Part of U.S. application Serial No. 07/465,863 (Attorney Docket No. 169.0011), filed January 16, 1990, which application is 5 a Continuation In Part of U.S. application Serial No. 07/405,499 (Attorney Docket No. 169.0007), filed September 11, 1989, which application is a Continuation In Part of U.S. application Serial No. 07/398,217 (Attorney Docket No. 169.0003), filed August 25, 1989, 10 which applications are entitled IMPROVED HLA TYPING METHOD AND REAGENTS THEREFOR by Malcolm J. Simons. Each of those applications is incorporated herein by reference in its entirety.

15 FIELD OF THE INVENTION

The present invention relates to a method for detection of alleles and haplotypes and reagents therefor.

20 BACKGROUND OF THE INVENTION

Due in part to a number of new analytical techniques, there has been a significant increase in knowledge about genetic information, particularly in humans. Allelic variants of genetic loci have been correlated to malignant and non-malignant monogenic and multigenic diseases. For example, monogenic diseases for which the defective gene has been identified include DuChenne muscular dystrophy, sickle-cell anemia, Lesch Nyhan syndrome, hemophilia, beta-thalassemia, cystic 25 fibrosis, polycystic kidney disease, ADA deficiency,  $\alpha$ -1-antitrypsin deficiency, Wilm's tumor and retinoblastoma. Other diseases which are believed to be monogenic for which the gene has not been identified include fragile X mental retardation and Huntington's 30 chorea.

Genes associated with multigenic diseases such as diabetes, colon cancer and premature coronary atherosclerosis have also been identified.

In addition to identifying individuals at risk for or carriers of genetic diseases, detection of allelic variants of a genetic locus has been used for organ transplantation, forensics, disputed paternity and a variety of other purposes in humans. In commercially important plants and animals, genes have not only been analyzed but genetically engineered and transmitted into other organisms.

A number of techniques have been employed to detect allelic variants of genetic loci including analysis of restriction fragment length polymorphic (RFLP) patterns, use of oligonucleotide probes, and DNA amplification methods. One of the most complicated groups of allelic variants, the major histocompatibility complex (MHC), has been extensively studied. The problems encountered in attempting to determine the HLA type of an individual are exemplary of problems encountered in characterizing other genetic loci.

The major histocompatibility complex is a cluster of genes that occupy a region on the short arm of chromosome 6. This complex, denoted the human leukocyte antigen (HLA) complex, includes at least 50 loci. For the purposes of HLA tissue typing, two main classes of loci are recognized. The Class I loci encode transplantation antigens and are designated A, B and C. The Class II loci (DRA, DRB, DQA1, DQB, DPA and DPB) encode products that control immune responsiveness. Of the Class II loci, all the loci are polymorphic with the exception of the DRA locus. That is, the DR $\alpha$  antigen polypeptide sequence is invariant.

HLA determinations are used in paternity determinations, transplant compatibility testing, forensics, blood component therapy, anthropological

studies, and in disease association correlations to diagnose disease or predict disease susceptibility. Due power of HLA to distinguish individuals and the need to match HLA type for transplantation, analytical methods to unambiguously characterize the alleles of the genetic loci associated with the complex have been sought. At present, DNA typing using RFLP and oligonucleotide probes has been used to type Class II locus alleles. Alleles of Class I loci and Class II DR and DQ loci are typically determined by serological methods. The alleles of the Class II DP locus are determined by primed lymphocyte typing (PLT).

Each of the HLA analysis methods has drawbacks. Serological methods require standard sera that are not widely available and must be continuously replenished. Additionally, serotyping is based on the reaction of the HLA gene products in the sample with the antibodies in the reagent sera. The antibodies recognize the expression products of the HLA genes on the surface of nucleated cells. The determination of fetal HLA type by serological methods may be difficult due to lack of maturation of expression of the antigens in fetal blood cells.

Oligonucleotide probe typing can be performed in two days and has been further improved by the recent use of polymerase chain reaction (PCR) amplification. PCR-based oligoprobe typing has been performed on Class II loci. Primed lymphocyte typing requires 5 to 10 days to complete and involves cell culture with its difficulties and inherent variability.

RFLP analysis is time consuming, requiring about 5 to 7 days to complete. Analysis of the fragment patterns is complex. Additionally, the technique requires the use of labelled probes. The most commonly used label,  $^{32}\text{P}$ , presents well known drawbacks associated with the use of radionuclides.

A fast, reliable method of genetic locus analysis is highly desirable.

#### DESCRIPTION OF THE PRIOR ART

5 U.S. Patent No. 4,683,195 (to Mullis et al, issued July 28, 1987) describes a process for amplifying, detecting and/or cloning nucleic acid sequences. The method involves treating separate complementary strands of DNA with two oligonucleotide primers, extending the  
10 primers to form complementary extension products that act as templates for synthesizing the desired nucleic acid sequence and detecting the amplified sequence. The method is commonly referred to as the polymerase chain reaction sequence amplification method or PCR.  
15 Variations of the method are described in U.S. Patent No. 4,683,194 (to Saiki et al, issued July 28, 1987). The polymerase chain reaction sequence amplification method is also described by Saiki et al, *Science*,  
230:1350-1354 (1985) and Scharf et al, *Science*, 324:163-  
20 166 (1986).

25 U.S. Patent No. 4,582,788 (to Erlich, issued April 15, 1986) describes an HLA typing method based on restriction length polymorphism (RFLP) and cDNA probes used therewith. The method is carried out by digesting an individual's HLA DNA with a restriction endonuclease that produces a polymorphic digestion pattern, subjecting the digest to genomic blotting using a labelled cDNA probe that is complementary to an HLA DNA sequence involved in the polymorphism, and comparing the resulting genomic blotting pattern with a standard.  
30 Locus-specific probes for Class II loci (DQ) are also described.

35 Kogan et al, *New Engl. J. Med.*, 317:985-990 (1987) describes an improved PCR sequence amplification method that uses a heat-stable polymerase (Taq polymerase) and high temperature amplification. The stringent

conditions used in the method provide sufficient fidelity of replication to permit analysis of the amplified DNA by determining DNA sequence lengths by visual inspection of an ethidium bromide-stained gel.

5 The method was used to analyze DNA associated with hemophilia A in which additional tandem repeats of a DNA sequence are associated with the disease and the amplified sequences were significantly longer than sequences that are not associated with the disease.

10 Simons and Erlich, pp 952-958 In: *Immunology of HLA* Vol. 1: Springer-Verlag, New York (1989) summarized RFLP-sequence interrelations at the DPA and DPB loci. RFLP fragment patterns analyzed with probes by Southern blotting provided distinctive patterns for DPw1-5 alleles and the corresponding DPB1 allele sequences, characterized two subtypic patterns for DPw2 and DPw4, and identified new DPw alleles.

15 20 Simons et al, pp 959-1023 In: *Immunology of HLA* Vol. 1: Springer-Verlag, New York (1989) summarized restriction length polymorphisms of HLA sequences for class II loci as determined by the 10th International Workshop Southern Blot Analysis. Southern blot analysis was shown to be suitable for typing of the major classes of HLA loci.

25 A series of three articles [Rommens et al, *Science* 245:1059-1065 (1989), Riordan et al, *Science* 245:1066-1072 (1989) and Kerem et al, *Science* 245:1073-1079 (1989) report a new gene analysis method called "jumping" used to identify the location of the CF gene, the sequence of the CF gene, and the defect in the gene and its percentage in the disease population, respectively.

30 35 DiLelia et al, *The Lancet* i:497-499 (1988) describes a screening method for detecting the two major alleles responsible for phenylketonuria in caucasians of Northern European descent. The mutations, located at

about the center of exon 12 and at the exon 12 junction with intervening sequence 12 are detected by PCR amplification of a 245 bp region of exon 12 and flanking intervening sequences. The amplified sequence encompasses both mutations and is analyzed using probes specific for each of the alleles (without prior electrophoretic separation).

5 Dicker et al, *BioTechniques* 7:830-837 (1989) and Mardis et al, *BioTechniques* 7:840-850 (1989) report on  
10 automated techniques for sequencing of DNA sequences, particularly PCR-generated sequences.

Each of the above-described references is incorporated herein by reference in its entirety.

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#### SUMMARY OF THE INVENTION

The present invention provides a method for detection of at least one allele of a genetic locus and can be used to provide direct determination of the haplotype. The method comprises amplifying genomic DNA with a primer pair that spans an intron sequence and defines a DNA sequence in genetic linkage with an allele to be detected. The primer-defined DNA sequence contains a sufficient number of intron sequence nucleotides to characterize the allele. Genomic DNA is amplified to produce an amplified DNA sequence characteristic of the allele. The amplified DNA sequence is analyzed to detect the presence of a genetic variation in the amplified DNA sequence such as a change in the length of the sequence, gain or loss of a restriction site or substitution of a nucleotide. The variation is characteristic of the allele to be detected.

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The present invention is based on the finding that intron sequences contain genetic variations that are characteristic of adjacent and remote alleles on the same chromosome. In particular, DNA sequences that

include a sufficient number of intron sequence nucleotides can be used for direct determination of haplotype.

The method can be used to detect alleles of genetic loci for any eukaryotic organism. Of particular interest are loci associated with malignant and nonmalignant monogenic and multigenic diseases, and identification of individual organisms or species in both plants and animals. In a preferred embodiment, the method is used to determine HLA allele type and haplotype.

Kits comprising one or more of the reagents used in the method are also described.

#### DETAILED DESCRIPTION OF THE INVENTION

The present invention provides a method for detection of alleles and haplotypes through analysis of intron sequence variation. The present invention is based on the discovery that amplification of intron sequences that exhibit linkage disequilibrium with adjacent and remote loci can be used to detect alleles of those loci. The present method reads haplotypes as the direct output of the intron typing analysis when a single, individual organism is tested. The method is particularly useful in humans but is generally applicable to all eukaryotes, and is preferably used to analyze plant and animal species.

The method comprises amplifying genomic DNA with a primer pair that spans an intron sequence and defines a DNA sequence in genetic linkage with an allele to be detected. Primer sites are located in conserved regions in the introns or exons bordering the intron sequence to be amplified. The primer-defined DNA sequence contains a sufficient number of intron sequence nucleotides to characterize the allele. The amplified DNA sequence is analyzed to detect the presence of a genetic variation

such as a change in the length of the sequence, gain or loss of a restriction site or substitution of a nucleotide.

5       The intron sequences provide genetic variations that, in addition to those found in exon sequences, further distinguish sample DNA, providing additional information about the individual organism. This information is particularly valuable for identification of individuals such as in paternity determinations and  
10      in forensic applications. The information is also valuable in any other application where heterozygotes (two different alleles) are to be distinguished from homozygotes (two copies of one allele).

15      More specifically, the present invention provides information regarding intron variation. Using the methods and reagents of this invention, two types of intron variation associated with genetic loci have been found. The first is allele-associated intron variation. That is, the intron variation pattern associates with the allele type at an adjacent locus. The second type of variation is associated with remote alleles (haplotypes). That is, the variation is present in individual organisms with the same genotype at the primary locus. Differences may occur between sequences of the same adjacent and remote locus types. However, individual-limited variation is uncommon.  
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30      Furthermore, an amplified DNA sequence that contains sufficient intron sequences will vary depending on the allele present in the sample. That is, the introns contain genetic variations (e.g. length polymorphisms due to insertions and/or deletions and changes in the number or location of restriction sites) which are associated with the particular allele of the locus and with the alleles at remote loci.  
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The reagents used in carrying out the methods of this invention are also described. The reagents can be

provided in kit form comprising one or more of the reagents used in the method.

#### *Definitions*

5       The term "allele", as used herein, means a genetic variation associated with a coding region; that is, an alternative form of the gene.

10      The term "linkage", as used herein, refers to the degree to which regions of genomic DNA are inherited together. Regions on different chromosomes do not exhibit linkage and are inherited together 50% of the time. Adjacent genes that are always inherited together exhibit 100% linkage.

15      The term "linkage disequilibrium", as used herein, refers to the co-occurrence of two alleles at linked loci such that the frequency of the co-occurrence of the alleles is greater than would be expected from the separate frequencies of occurrence of each allele. Alleles that co-occur with frequencies expected from their separate frequencies are said to be in "linkage equilibrium".

20      As used herein, "haplotype" is a region of genomic DNA on a chromosome which is bounded by recombination sites such that genetic loci within a haplotypic region are usually inherited as a unit. However, occasionally, genetic rearrangements may occur within a haplotype. Thus, the term haplotype is an operational term that refers to the occurrence on a chromosome of linked loci.

25      As used herein, the term "intron" refers to untranslated DNA sequences between exons, together with 5' and 3' untranslated regions associated with a genetic locus. In addition, the term is used to refer to the spacing sequences between genetic loci (intergenic spacing sequences) which are not associated with a coding region and are colloquially referred to as "junk". While the art traditionally uses the term

"intron" to refer only to untranslated sequences between exons, this expanded definition was necessitated by the lack of any art recognized term which encompasses all non-exon sequences.

5 As used herein, an "intervening sequence" is an intron which is located between two exons within a gene. The term does not encompass upstream and downstream noncoding sequences associated with the genetic locus.

10 As used herein, the term "amplified DNA sequence" refers to DNA sequences which are copies of a portion of a DNA sequence and its complementary sequence, which copies correspond in nucleotide sequence to the original DNA sequence and its complementary sequence.

15 The term "complement", as used herein, refers to a DNA sequence that is complementary to a specified DNA sequence.

The term "primer site", as used herein, refers to the area of the target DNA to which a primer hybridizes.

20 The term "primer pair", as used herein, means a set of primers including a 5' upstream primer that hybridizes with the 5' end of the DNA sequence to be amplified and a 3' downstream primer that hybridizes with the complement of the 3' end of the sequence to be amplified.

25 The term "exon-limited primers", as used herein, means a primer pair having primers located within or just outside of an exon in a conserved portion of the intron, which primers amplify a DNA sequence which includes an exon or a portion thereof and not more than a small, para-exon region of the adjacent intron(s).

30 The term "intron-spanning primers", as used herein, means a primer pair that amplifies at least a portion of one intron, which amplified intron region includes sequences which are not conserved. The intron-spanning primers can be located in conserved regions of

the introns or in adjacent, upstream and/or downstream exon sequences.

5           The term "genetic locus", as used herein, means the region of the genomic DNA that includes the gene that encodes a protein including any upstream or downstream transcribed noncoding regions and associated regulatory regions. Therefore, an HLA locus is the region of the genomic DNA that includes the gene that encodes an HLA gene product.

10          As used herein, the term "adjacent locus" refers to either (1) the locus in which a DNA sequence is located or (2) the nearest upstream or downstream genetic locus for intron DNA sequences not associated with a genetic locus.

15          As used herein, the term "remote locus" refers to either (1) a locus which is upstream or downstream from the locus in which a DNA sequence is located or (2) for intron sequences not associated with a genetic locus, a locus which is upstream or downstream from the nearest upstream or downstream genetic locus to the intron sequence.

20          As used herein, the term "locus-specific primer", as used herein, means a primer that specifically hybridizes with a portion of the stated gene locus or its complementary strand, at least for one allele of the locus, and does not hybridize with other DNA sequences under the conditions used in the amplification method.

25          As used herein, the terms "endonuclease" and "restriction endonuclease" refer to an enzyme that cuts double-stranded DNA having a particular nucleotide sequence. The specificities of numerous endonucleases are well known and can be found in a variety of publications, e.g. *Molecular Cloning: A Laboratory Manual* by Maniatis et al, Cold Spring Harbor Laboratory 1982. That manual is incorporated herein by reference in its entirety.

The term "restriction fragment length polymorphism" (or RFLP), as used herein, refers to differences in DNA nucleotide sequences that produce fragments of different lengths when cleaved by a restriction endonuclease.

5           The term "primer-defined length polymorphisms" (or PDLP), as used herein, refers to differences in the lengths of amplified DNA sequences due to insertions or deletions in the intron region of the locus included in  
10          the amplified DNA sequence.

10          The term "HLA DNA", as used herein, means DNA that includes the genes that encode HLA antigens. HLA DNA is found in all nucleated human cells.

15          *Primers*  
15          The method of this invention is based on amplification of selected intron regions of genomic DNA. The methodology is facilitated by the use of primers that selectively amplify DNA associated with one or more alleles of a genetic locus of interest and not with other genetic loci.

20          25          A locus-specific primer pair contains a 5' upstream primer that defines the 5' end of the amplified sequence by hybridizing with the 5' end of the target sequence to be amplified and a 3' downstream primer that defines the 3' end of the amplified sequence by hybridizing with the complement of the 3' end of the DNA sequence to be amplified. The primers in the primer pair do not hybridize with DNA of other genetic loci under the conditions used in the present invention.

30          35          For each primer of the locus-specific primer pair, the primer hybridizes to at least one allele of the DNA locus to be amplified or to its complement. A primer pair can be prepared for each allele of a selected locus, which primer pair amplifies only DNA for the selected locus. In this way combinations of primer

5 pairs can be used to amplify genomic DNA of a particular locus, irrespective of which allele is present in a sample. Preferably, the primer pair amplifies DNA of at least two, more preferably more than two, alleles of a locus. In a most preferred embodiment, the primer sites are conserved, and thus amplify all haplotypes.

10 However, primer pairs or combinations thereof that specifically bind with the most common alleles present in a particular population group are also contemplated.

15 The amplified DNA sequence that is defined by the primers contains a sufficient number of intron sequence nucleotides to distinguish between at least two alleles of an adjacent locus, and preferably, to identify the allele of the locus which is present in the sample. For some purposes, the sequence can also be selected to contain sufficient genetic variations to distinguish between individual organisms with the same allele or to distinguish between haplotypes.

20 Length of sequence

25 The length of the amplified sequence which is required to include sufficient genetic variability to enable discrimination between all alleles of a locus bears a direct relation to the extent of the polymorphism of the locus (the number of alleles). That is, as the number of alleles of the tested locus increases, the size of an amplified sequence which contains sufficient genetic variations to identify each allele increases. For a particular population group, 30 one or more of the recognized alleles for any given locus may be absent from that group and need not be considered in determining a sequence which includes sufficient variability for that group. Conveniently, however, the primer pairs are selected to amplify a DNA sequence which is sufficient to distinguish between all

recognized alleles of the tested locus. The same considerations apply when a haplotype is determined.

For example, the least polymorphic HLA locus is DPA which currently has four recognized alleles. For that locus, a primer pair which amplifies only a portion of the variable exon encoding the allelic variation contains sufficient genetic variability to distinguish between the alleles when the primer sites are located in an appropriate region of the variable exon. Exon-limited primers can be used to produce an amplified sequence that includes as few as about 200 nucleotides (nt). However, as the number of alleles of the locus increases, the number of genetic variations in the sequence must increase to distinguish all alleles.

Addition of invariant exon sequences provides no additional genetic variation. When about eight or more alleles are to be distinguished, as for the DQA1 locus and more variable loci, amplified sequences should extend into at least one intron in the locus, preferably an intron adjacent to the variable exon.

Additionally, where alleles of the locus exist which differ by a single basepair in the variable exon, intron sequences are included in amplified sequences to provide sufficient variability to distinguish alleles. For example, for the DQA1 locus (with eight currently recognized alleles) and the DPB locus (with 24 alleles), the DQA1.1/1.2 (now referred to as DQA1 0101/0102) and DPB2.1/4.2 (now referred to as DPB0201/0402) alleles differ by a single basepair. To distinguish those alleles, amplified sequences which include an intron sequence region are required. About 300 to 500 nucleotides is sufficient, depending on the location of the sequence. That is, 300 to 500 nucleotides comprised primarily of intron sequence nucleotides sufficiently close to the variable exon are sufficient.

For loci with more extensive polymorphisms (such as DQB with 14 currently recognized alleles, DPB with 24 currently recognized alleles, DRB with 34 currently recognized alleles and for each of the Class I loci), the amplified sequences need to be larger to provide sufficient variability to distinguish between all the alleles. An amplified sequence that includes at least about 0.5 kilobases (Kb), preferably at least about 1.0 Kb, more preferably at least about 1.5 Kb generally provides a sufficient number of restriction sites for loci with extensive polymorphisms. The amplified sequences used to characterize highly polymorphic loci are generally between about 800 to about 2,000 nucleotides (nt), preferably between about 1000 to about 1800 nucleotides in length.

When haplotype information regarding remote alleles is desired, the sequences are generally between about 1,000 to about 2,000 nt in length. Longer sequences are required when the amplified sequence encompasses highly conserved regions such as exons or highly conserved intron regions, e.g., promoters, operators and other DNA regulatory regions. Longer amplified sequences (including more intron nucleotide sequences) are also required as the distance between the amplified sequences and the allele to be detected increases.

Highly conserved regions included in the amplified DNA sequence, such as exon sequences or highly conserved intron sequences (e.g. promoters, enhancers, or other regulatory regions) may provide little or no genetic variation. Therefore, such regions do not contribute, or contribute only minimally, to the genetic variations present in the amplified DNA sequence. When such regions are included in the amplified DNA sequence, additional nucleotides may be required to encompass sufficient genetic variations to distinguish alleles, in

comparison to an amplified DNA sequence of the same length including only intron sequences.

Location of the amplified DNA sequence

5       The amplified DNA sequence is located in a region of genomic DNA that contains genetic variation which is in genetic linkage with the allele to be detected.

10      Preferably, the sequence is located in an intron sequence adjacent to an exon of the genetic locus. More

15      preferably, the amplified sequence includes an intervening sequence adjacent to an exon that encodes the allelic variability associated with the locus (a variable exon). The sequence preferably includes at least a portion of one of the introns adjacent to a

20      variable exon and can include a portion of the variable exon. When additional sequence information is required, the amplified DNA sequence preferably encompasses a variable exon and all or a portion of both adjacent intron sequences.

25      Alternatively, the amplified sequence can be in an intron which does not border an exon of the genetic locus. Such introns are located in the downstream or upstream gene flanking regions or even in an intervening sequence in another genetic locus which is in linkage disequilibrium with the allele to be detected.

30      For some genetic loci, genomic DNA sequences may not be available. When only cDNA sequences are available and intron locations within the sequence are not identified, primers are selected at intervals of about 200 nt and used to amplify genomic DNA. If the amplified sequence contains about 200 nt, the location of the first primer is moved about 200 nt to one side of the second primer location and the amplification is repeated until either (1) an amplified DNA sequence that is larger than expected is produced or (2) no amplified DNA sequence is produced. In either case, the location

of an intron sequence has been determined. The same methodology can be used when only the sequence of a marker site that is highly linked to the genetic locus is available, as is the case for many genes associated with inherited diseases.

When the amplified DNA sequence does not include all or a portion of an intron adjacent to the variable exon(s), the sequence must also satisfy a second requirement. The amplified sequence must be sufficiently close to the variable exon(s) to exclude recombination and loss of linkage disequilibrium between the amplified sequence and the variable exon(s). This requirement is satisfied if the regions of the genomic DNA are within about 5 Kb, preferably within about 4 Kb, most preferably within 2 Kb of the variable exon(s). The amplified sequence can be outside of the genetic locus but is preferably within the genetic locus.

Preferably, for each primer pair, the amplified DNA sequence defined by the primers includes at least 200 nucleotides, and more preferably at least 400 nucleotides, of an intervening sequence adjacent to the variable exon(s). Although the variable exon usually provides fewer variations in a given number of nucleotides than an adjacent intervening sequence, each of those variations provides allele-relevant information. Therefore, inclusion of the variable exon provides an advantage.

Since PCR methodology can be used to amplify sequences of several Kb, the primers can be located so that additional exons or intervening sequences are included in the amplified sequence. Of course, the increased size of the amplified DNA sequence increases the chance of replication error, so addition of invariant regions provides some disadvantages. However, those disadvantages are not as likely to affect an analysis based on the length of the sequence or the RFLP

fragment patterns as one based on sequencing the amplification product. For particular alleles, especially those with highly similar exon sequences, amplified sequences of greater than about 1 or 1.5 Kb 5 may be necessary to discriminate between all alleles of a particular locus.

The ends of the amplified DNA sequence are defined by the primer pair used in the amplification. Each 10 primer sequence must correspond to a conserved region of the genomic DNA sequence. Therefore, the location of the amplified sequence will, to some extent, be dictated by the need to locate the primers in conserved regions. When sufficient intron sequence information to determine 15 conserved intron regions is not available, the primers can be located in conserved portions of the exons and used to amplify intron sequences between those exons.

When appropriately-located, conserved sequences are not unique to the genetic locus, a second primer 20 located within the amplified sequence produced by the first primer pair can be used to provide an amplified DNA sequence specific for the genetic locus. At least one of the primers of the second primer pair is located 25 in a conserved region of the amplified DNA sequence defined by the first primer pair. The second primer pair is used following amplification with the first primer pair to amplify a portion of the amplified DNA sequence produced by the first primer pair.

There are three major types of genetic variations 30 that can be detected and used to identify an allele. Those variations, in order of ease of detection, are 35 (1) a change in the length of the sequence, (2) a change in the presence or location of at least one restriction site and (3) the substitution of one or a few nucleotides that does not result in a change in a restriction site. Other variations within the amplified DNA sequence are also detectable.

There are three types of techniques which can be used to detect the variations. The first is sequencing the amplified DNA sequence. Sequencing is the most time consuming and also the most revealing analytical method, since it detects any type of genetic variation in the amplified sequence. The second analytical method uses allele-specific oligonucleotide or sequence-specific oligonucleotides probes (ASO or SSO probes). Probes can detect single nucleotide changes which result in any of the types of genetic variations, so long as the exact sequence of the variable site is known. A third type of analytical method detects sequences of different lengths (e.g., due to an insertion or deletion or a change in the location of a restriction site) and/or different numbers of sequences (due to either gain or loss of restriction sites). A preferred detection method is by gel or capillary electrophoresis. To detect changes in the lengths of fragments or the number of fragments due to changes in restriction sites, the amplified sequence must be digested with an appropriate restriction endonuclease prior to analysis of fragment length patterns.

The first genetic variation is a difference in the length of the primer-defined amplified DNA sequence, referred to herein as a primer-defined length polymorphism (PDLP), which difference in length distinguishes between at least two alleles of the genetic locus. The PDLPs result from insertions or deletions of large stretches (in comparison to the total length of the amplified DNA sequence) of DNA in the portion of the intron sequence defined by the primer pair. To detect PDLPs, the amplified DNA sequence is located in a region containing insertions or deletions of a size that is detectable by the chosen method. The amplified DNA sequence should have a length which provides optimal resolution of length differences. For

electrophoresis, DNA sequences of about 300 to 500 bases  
in length provide optimal resolution of length  
differences. Nucleotide sequences which differ in  
length by as few as 3 nt, preferably 25 to 50 nt, can be  
5 distinguished. However, sequences as long as 800 to  
2,000 nt which differ by at least about 50 nt are also  
readily distinguishable. Gel electrophoresis and  
capillary electrophoresis have similar limits of  
resolution. Preferably the length differences between  
10 amplified DNA sequences will be at least 10, more  
preferably 20, most preferably 50 or more, nt between  
the alleles. Preferably, the amplified DNA sequence is  
between 300 to 1,000 nt and encompasses length  
differences of at least 3, preferably 10 or more nt.

15 Preferably, the amplified sequence is located in  
an area which provides PDLP sequences that distinguish  
most or all of the alleles of a locus. An example of  
PDLP-based identification of five of the eight DQ<sub>A1</sub>  
alleles is described in detail in the examples.

20 When the variation to be detected is a change in a  
restriction site, the amplified DNA sequence necessarily  
contains at least one restriction site which (1) is  
present in one allele and not in another, (2) is  
apparently located in a different position in the  
sequence of at least two alleles, or (3) combinations  
25 thereof. The amplified sequence will preferably be  
located such that restriction endonuclease cleavage  
produces fragments of detectably different lengths,  
rather than two or more fragments of approximately the  
same length.

30 For allelic differences detected by ASO or SSO  
probes, the amplified DNA sequence includes a region of  
from about 200 to about 400 nt which is present in one  
or more alleles and not present in one or more other  
alleles. In a most preferred embodiment, the sequence  
35 contains a region detectable by a probe that is present

in only one allele of the genetic locus. However, combinations of probes which react with some alleles and not others can be used to characterize the alleles.

For the method described herein, it is contemplated that use of more than one amplified DNA sequence and/or use of more than one analytical method per amplified DNA sequence may be required for highly polymorphic loci, particularly for loci where alleles differ by single nucleotide substitutions that are not unique to the allele or when information regarding remote alleles (haplotypes) is desired. More particularly, it may be necessary to combine a PDLP analysis with an RFLP analysis, to use two or more amplified DNA sequences located in different positions or to digest a single amplified DNA sequence with a plurality of endonucleases to distinguish all the alleles of some loci. These combinations are intended to be included within the scope of this invention.

For example, the analysis of the haplotypes of DQA1 locus described in the examples uses PDLPs and RFLP analysis using three different enzyme digests to distinguish the eight alleles and 20 of the 32 haplotypes of the locus.

25 Length and sequence homology of primers

Each locus-specific primer includes a number of nucleotides which, under the conditions used in the hybridization, are sufficient to hybridize with an allele of the locus to be amplified and to be free from hybridization with alleles of other loci. The specificity of the primer increases with the number of nucleotides in its sequence under conditions that provide the same stringency. Therefore, longer primers are desirable. Sequences with fewer than 15 nucleotides are less certain to be specific for a particular locus. That is, sequences with fewer than 15 nucleotides are

more likely to be present in a portion of the DNA associated with other genetic loci, particularly loci of other common origin or evolutionarily closely related origin, in inverse proportion to the length of the nucleotide sequence.

5           Each primer preferably includes at least about 15 nucleotides, more preferably at least about 20 nucleotides. The primer preferably does not exceed about 30 nucleotides, more preferably about 25

10          nucleotides. Most preferably, the primers have between about 20 and about 25 nucleotides.

15          A number of preferred primers are described herein. Each of those primers hybridizes with at least about 15 consecutive nucleotides of the designated region of the allele sequence. For many of the primers, the sequence is not identical for all of the other alleles of the locus. For each of the primers, additional preferred primers have sequences which correspond to the sequences of the homologous region of other alleles of the locus or to their complements.

20          When two sets of primer pairs are used sequentially, with the second primer pair amplifying the product of the first primer pair, the primers can be the same size as those used for the first amplification. However, smaller primers can be used in the second amplification and provide the requisite specificity. These smaller primers can be selected to be allele-specific, if desired. The primers of the second primer pair can have 15 or fewer, preferably 8 to 12, more preferably 8 to 10 nucleotides. When two sets of primer pairs are used to produce two amplified sequences, the second amplified DNA sequence is used in the subsequent analysis of genetic variation and must meet the requirements discussed previously for the amplified DNA sequence.

The primers preferably have a nucleotide sequence  
that is identical to a portion of the DNA sequence to be  
amplified or its complement. However, a primer having  
two nucleotides that differ from the target DNA sequence  
5 or its complement also can be used. Any nucleotides  
that are not identical to the sequence or its complement  
are preferably not located at the 3' end of the primer.  
The 3' end of the primer preferably has at least two,  
preferably three or more, nucleotides that are  
10 complementary to the sequence to which the primer binds.  
Any nucleotides that are not identical to the sequence  
to be amplified or its complement will preferably not be  
adjacent in the primer sequence. More preferably,  
noncomplementary nucleotides in the primer sequence will  
15 be separated by at least three, more preferably at least  
five, nucleotides. The primers should have a melting  
temperature ( $T_m$ ) from about 55 to 75°C. Preferably the  
 $T_m$  is from about 60°C to about 65°C to facilitate  
stringent amplification conditions.

20 The primers can be prepared using a number of  
methods, such as, for example, the phosphotriester and  
phosphodiester methods or automated embodiments thereof.  
The phosphodiester and phosphotriester methods are  
described in Cruthers, *Science* 230:281-285 (1985); Brown  
25 et al, *Meth. Enzymol.*, 68:109 (1979); and Nrang et al,  
*Meth. Enzymol.*, 68:90 (1979). In one automated method,  
diethylphosphoramidites which can be synthesized as  
described by Beaucage et al, *Tetrahedron letters*,  
22:1859-1962 (1981) are used as starting materials. A  
30 method for synthesizing primer oligonucleotide sequences  
on a modified solid support is described in U.S. Pat.  
No. 4,458,066. Each of the above references is  
incorporated herein by reference in its entirety.

Exemplary primer sequences for analysis of Class I  
35 and Class II HLA loci; bovine leukocyte antigens, and  
cystic fibrosis are described herein.

### Amplification

The locus-specific primers are used in an amplification process to produce a sufficient amount of DNA for the analysis method. For production of RFLP fragment patterns or PDLP patterns which are analyzed by electrophoresis, about 1 to about 500 ng of DNA is required. A preferred amplification method is the polymerase chain reaction (PCR). PCR amplification methods are described in U.S. Patent No. 4,683,195 (to Mullis et al, issued July 28, 1987); U.S Patent No. 4,683,194 (to Saiki et al, issued July 28, 1987); Saiki et al, *Science*, 230:1350-1354 (1985); Scharf et al, *Science*, 324:163-166 (1986); Kogan et al, *New Engl. J. Med.*, 317:985-990 (1987) and Saiki, Gyllensten and Erlich, *The Polymerase Chain Reaction in Genome Analysis: A Practical Approach*, ed. Davies pp. 141-152, (1988) I.R.L. Press, Oxford. Each of the above references is incorporated herein by reference in its entirety.

Prior to amplification, a sample of the individual organism's DNA is obtained. All nucleated cells contain genomic DNA and, therefore, are potential sources of the required DNA. For higher animals, peripheral blood cells are typically used rather than tissue samples. As little as 0.01 to 0.05 cc of peripheral blood provides sufficient DNA for amplification. Hair, semen and tissue can also be used as samples. In the case of fetal analyses, placental cells or fetal cells present in amniotic fluid can be used. The DNA is isolated from nucleated cells under conditions that minimize DNA degradation. Typically, the isolation involves digesting the cells with a protease that does not attack DNA at a temperature and pH that reduces the likelihood of DNase activity. For peripheral blood cells, lysing

the cells with a hypotonic solution (water) is sufficient to release the DNA.

DNA isolation from nucleated cells is described by Kan et al, *N. Engl. J. Med.* **297**:1080-1084 (1977); Kan et al, *Nature* **251**:392-392 (1974); and Kan et al, *PNAS* **75**:5631-5635 (1978). Each of the above references is incorporated herein by reference in its entirety.

Extraction procedures for samples such as blood, semen, hair follicles, semen, mucous membrane epithelium and other sources of genomic DNA are well known. For plant cells, digestion of the cells with cellulase releases DNA. Thereafter DNA is purified as described above.

The extracted DNA can be purified by dialysis, chromatography, or other known methods for purifying polynucleotides prior to amplification. Typically, the DNA is not purified prior to amplification.

The amplified DNA sequence is produced by using the portion of the DNA and its complement bounded by the primer pair as a template. As a first step in the method, the DNA strands are separated into single stranded DNA. This strand separation can be accomplished by a number of methods including physical or chemical means. A preferred method is the physical method of separating the strands by heating the DNA until it is substantially (approximately 93%) denatured. Heat denaturation involves temperatures ranging from about 80° to 105°C for times ranging from about 15 to 30 seconds. Typically, heating the DNA to a temperature of from 90° to 93°C for about 30 seconds to about 1 minute is sufficient.

The primer extension product(s) produced are complementary to the primer-defined region of the DNA and hybridize therewith to form a duplex of equal length strands. The duplexes of the extension products and their templates are then separated into single-stranded DNA. When the complementary strands of the duplexes are

separated, the strands are ready to be used as a template for the next cycle of synthesis of additional DNA strands.

Each of the synthesis steps can be performed using conditions suitable for DNA amplification. Generally, the amplification step is performed in a buffered aqueous solution, preferably at a pH of about 7 to about 9, more preferably about pH 8. A suitable amplification buffer contains Tris-HCl as a buffering agent in the range of about 10 to 100 mM. The buffer also includes a monovalent salt, preferably at a concentration of at least about 10 mM and not greater than about 60 mM. Preferred monovalent salts are KCl, NaCl and  $(\text{NH}_4)_2\text{SO}_4$ . The buffer also contains  $\text{MgCl}_2$  at about 5 to 50 mM. Other buffering systems such as hepes or glycine-NaOH and potassium phosphate buffers can be used. Typically, the total volume of the amplification reaction mixture is about 50 to 100  $\mu\text{l}$ .

Preferably, for genomic DNA, a molar excess of about  $10^6:1$  primer:template of the primer pair is added to the buffer containing the separated DNA template strands. A large molar excess of the primers improves the efficiency of the amplification process. In general, about 100 to 150 ng of each primer is added.

The deoxyribonucleotide triphosphates dATP, dCTP, dGTP and dTTP are also added to the amplification mixture in amounts sufficient to produce the amplified DNA sequences. Preferably, the dNTPs are present at a concentration of about 0.75 to about 4.0 mM, more preferably about 2.0 mM. The resulting solution is heated to about 90° to 93°C for from about 30 seconds to about 1 minute to separate the strands of the DNA. After this heating period the solution is cooled to the amplification temperature.

Following separation of the DNA strands, the primers are allowed to anneal to the strands. The

annealing temperature varies with the length and GC content of the primers. Those variables are reflected in the  $T_m$  of each primer. Exemplary HLA DQA1 primers of this invention, described below, require temperatures of about 55°C. The exemplary HLA Class I primers of this invention require slightly higher temperatures of about 62° to about 68°C. The extension reaction step is performed following annealing of the primers to the genomic DNA.

An appropriate agent for inducing or catalyzing the primer extension reaction is added to the amplification mixture either before or after the strand separation (denaturation) step, depending on the stability of the agent under the denaturation conditions. The DNA synthesis reaction is allowed to occur under conditions which are well known in the art. This synthesis reaction (primer extension) can occur at from room temperature up to a temperature above which the polymerase no longer functions efficiently. Elevating the amplification temperature enhances the stringency of the reaction. As stated previously, stringent conditions are necessary to ensure that the amplified sequence and the DNA template sequence contain the same nucleotide sequence, since substitution of nucleotides can alter the restriction sites or probe binding sites in the amplified sequence.

The inducing agent may be any compound or system which facilitates synthesis of primer extension products, preferably enzymes. Suitable enzymes for this purpose include DNA polymerases (such as, for example, E. coli DNA polymerase I, Klenow fragment of E. coli DNA polymerase I, T4 DNA polymerase), reverse transcriptase, and other enzymes (including heat-stable polymerases) which facilitate combination of the nucleotides in the proper manner to form the primer extension products. Most preferred is Taq polymerase or other heat-stable

polymerases which facilitate DNA synthesis at elevated temperatures (about 60° to 90°C). Taq polymerase is described, e.g., by Chien et al, *J. Bacteriol.*, 127:1550-1557 (1976). That article is incorporated herein by reference in its entirety. When the extension step is performed at about 72°C, about 1 minute is required for every 1000 bases of target DNA to be amplified.

The synthesis of the amplified sequence is initiated at the 3' end of each primer and proceeds toward the 5' end of the template along the template DNA strand, until synthesis terminates, producing DNA sequences of different lengths. The newly synthesized strand and its complementary strand form a double-stranded molecule which is used in the succeeding steps of the process. In the next step, the strands of the double-stranded molecule are separated (denatured) as described above to provide single-stranded molecules.

New DNA is synthesized on the single-stranded template molecules. Additional polymerase, nucleotides and primers can be added if necessary for the reaction to proceed under the conditions described above. After this step, half of the extension product consists of the amplified sequence bounded by the two primers. The steps of strand separation and extension product synthesis can be repeated as many times as needed to produce the desired quantity of the amplified DNA sequence. The amount of the amplified sequence produced accumulates exponentially. Typically, about 25 to 30 cycles are sufficient to produce a suitable amount of the amplified DNA sequence for analysis.

The amplification method can be performed in a step-wise fashion where after each step new reagents are added, or simultaneously, where all reagents are added at the initial step, or partially step-wise and partially simultaneously, where fresh reagent is added

after a given number of steps. The amplification reaction mixture can contain, in addition to the sample genomic DNA, the four nucleotides, the primer pair in molar excess, and the inducing agent, e.g., Tag polymerase.

5            Each step of the process occurs sequentially notwithstanding the initial presence of all the reagents. Additional materials may be added as necessary. Typically, the polymerase is not replenished  
10          when using a heat-stable polymerase. After the appropriate number of cycles to produce the desired amount of the amplified sequence, the reaction may be halted by inactivating the enzymes, separating the components of the reaction or stopping the thermal  
15          cycling.

20          In a preferred embodiment of the method, the amplification includes the use of a second primer pair to perform a second amplification following the first amplification. The second primer pair defines a DNA sequence which is a portion of the first amplified sequence. That is, at least one of the primers of the second primer pair defines one end of the second amplified sequence which is within the ends of the first amplified sequence. In this way, the use of the second primer pair helps to ensure that any amplified sequence produced in the second amplification reaction is specific for the tested locus. That is, non-target sequences which may be copied by a locus-specific pair are unlikely to contain sequences that hybridize with a second locus-specific primer pair located within the first amplified sequence.  
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In another embodiment, the second primer pair is specific for one allele of the locus. In this way, detection of the presence of a second amplified sequence indicates that the allele is present in the sample. The presence of a second amplified sequence can be

determined by quantitating the amount of DNA at the start and the end of the second amplification reaction. Methods for quantitating DNA are well known and include determining the optical density at 260 ( $OD_{260}$ ), and preferably additionally determining the ratio of the optical density at 260 to the optical density at 280 ( $OD_{260}/OD_{280}$ ) to determine the amount of DNA in comparison to protein in the sample.

Preferably, the first amplification will contain sufficient primer for only a limited number of primer extension cycles, e.g. less than 15, preferably about 10 to 12 cycles, so that the amount of amplified sequence produced by the process is sufficient for the second amplification but does not interfere with a determination of whether amplification occurred with the second primer pair. Alternatively, the amplification reaction can be continued for additional cycles and aliquoted to provide appropriate amounts of DNA for one or more second amplification reactions. Approximately 100 to 150 ng of each primer of the second primer pair is added to the amplification reaction mixture. The second set of primers is preferably added following the initial cycles with the first primer pair. The amount of the first primer pair can be limited in comparison to the second primer pair so that, following addition of the second pair, substantially all of the amplified sequences will be produced by the second pair.

As stated previously, the DNA can be quantitated to determine whether an amplified sequence was produced in the second amplification. If protein in the reaction mixture interferes with the quantitation (usually due to the presence of the polymerase), the reaction mixture can be purified, as by using a 100,000 MW cut off filter. Such filters are commercially available from Millipore and from Centricon.

Analysis of the Amplified DNA Sequence

As discussed previously, the method used to analyze the amplified DNA sequence to characterize the allele(s) present in the sample DNA depends on the genetic variation in the sequence. When distinctions between alleles include primer-defined length polymorphisms, the amplified sequences are separated based on length, preferably using gel or capillary electrophoresis. When using probe hybridization for analysis, the amplified sequences are reacted with labeled probes. When the analysis is based on RFLP fragment patterns, the amplified sequences are digested with one or more restriction endonucleases to produce a digest and the resultant fragments are separated based on length, preferably using gel or capillary electrophoresis. When the only variation encompassed by the amplified sequence is a sequence variation that does not result in a change in length or a change in a restriction site and is unsuitable for detection by a probe, the amplified DNA sequences are sequenced.

Procedures for each step of the various analytical methods are well known and are described below.

*Production of RFLP Fragment Patterns*

Restriction endonucleases

A restriction endonuclease is an enzyme that cleaves or cuts DNA hydrolytically at a specific nucleotide sequence called a restriction site. Endonucleases that produce blunt end DNA fragments (hydrolysis of the phosphodiester bonds on both DNA strands occur at the same site) as well as endonucleases that produce sticky ended fragments (the hydrolysis sites on the strands are separated by a few nucleotides from each other) can be used.

Restriction enzymes are available commercially from a number of sources including Sigma

Pharmaceuticals, Bethesda Research Labs, Boehringer-Manheim and Pharmacia. As stated previously, a restriction endonuclease used in the present invention cleaves an amplified DNA sequence of this invention to produce a digest comprising a set of fragments having distinctive fragment lengths. In particular, the fragments for one allele of a locus differ in size from the fragments for other alleles of the locus. The patterns produced by separation and visualization of the fragments of a plurality of digests are sufficient to distinguish each allele of the locus. More particularly, the endonucleases are chosen so that by using a plurality of digests of the amplified sequence, preferably fewer than five, more preferably two or three digests, the alleles of a locus can be distinguished.

In selecting an endonuclease, the important consideration is the number of fragments produced for amplified sequences of the various alleles of a locus. More particularly, a sufficient number of fragments must be produced to distinguish between the alleles and, if required, to provide for individuality determinations. However, the number of fragments must not be so large or so similar in size that a pattern that is not distinguishable from those of other haplotypes by the particular detection method is produced. Preferably, the fragments are of distinctive sizes for each allele. That is, for each endonuclease digest of a particular amplified sequence, the fragments for an allele preferably differ from the fragments for every other allele of the locus by at least 10, preferably 20, more preferably 30, most preferably 50 or more nucleotides.

One of ordinary skill can readily determine whether an endonuclease produces RFLP fragments having distinctive fragment lengths. The determination can be made experimentally by cleaving an amplified sequence for each allele with the designated endonuclease in the

invention method. The fragment patterns can then be analyzed. Distinguishable patterns will be readily recognized by determining whether comparison of two or more digest patterns is sufficient to demonstrate characteristic differences between the patterns of the alleles.

The number of digests that need to be prepared for any particular analysis will depend on the desired information and the particular sample to be analyzed. Since HLA analyses are used for a variety of purposes ranging from individuality determinations for forensics and paternity to tissue typing for transplantation, the HLA complex will be used as exemplary.

A single digest may be sufficient to determine that an individual cannot be the person whose blood was found at a crime scene. In general, however, where the DNA samples do not differ, the use of two to three digests for each of two to three HLA loci will be sufficient for matching applications (forensics, paternity). For complete HLA typing, each locus needs to be determined.

In a preferred embodiment, sample HLA DNA sequences are divided into aliquots containing similar amounts of DNA per aliquot and are amplified with primer pairs (or combinations of primer pairs) to produce amplified DNA sequences for a number of HLA loci. Each amplification mixture contains only primer pairs for one HLA locus. The amplified sequences are preferably processed concurrently, so that a number of digest RFLP fragment patterns can be produced from one sample. In this way, the HLA type for a number of alleles can be determined simultaneously.

Alternatively, preparation of a number of RFLP fragment patterns provides additional comparisons of patterns to distinguish samples for forensic and paternity analyses where analysis of one locus

frequently fails to provide sufficient information for the determination when the sample DNA has the same allele as the DNA to which it is compared.

5       Production of RFLP fragments

Following amplification, the amplified DNA sequence is combined with an endonuclease that cleaves or cuts the amplified DNA sequence hydrolytically at a specific restriction site. The combination of the endonuclease with the amplified DNA sequence produces a digest containing a set of fragments having distinctive fragment lengths. U.S. Patent No. 4,582,788 (to Erlich, issued April 15, 1986) describes an HLA typing method based on restriction length polymorphism (RFLP). That patent is incorporated herein by reference in its entirety.

In a preferred embodiment, two or more aliquots of the amplification reaction mixture having approximately equal amounts of DNA per aliquot are prepared. Conveniently about 5 to about 10  $\mu$ l of a 100  $\mu$ l reaction mixture is used for each aliquot. Each aliquot is combined with a different endonuclease to produce a plurality of digests. In this way, by using a number of endonucleases for a particular amplified DNA sequence, locus-specific combinations of endonucleases that distinguish a plurality of alleles of a particular locus can be readily determined. Following preparation of the digests, each of the digests can be used to form RFLP patterns. Preferably, two or more digests can be pooled prior to pattern formation.

Alternatively, two or more restriction endonucleases can be used to produce a single digest. The digest differs from one where each enzyme is used separately and the resultant fragments are pooled since fragments produced by one enzyme may include one or more restriction sites recognized by another enzyme in the

digest. Patterns produced by simultaneous digestion by two or more enzymes will include more fragments than pooled products of separate digestions using those enzymes and will be more complex to analyze.

5 Furthermore, one or more restriction endonucleases can be used to digest two or more amplified DNA sequences. That is, for more complete resolution of all the alleles of a locus, it may be desirable to produce 10 amplified DNA sequences encompassing two different regions. The amplified DNA sequences can be combined and digested with at least one restriction endonuclease to produce RFLP patterns.

15 The digestion of the amplified DNA sequence with the endonuclease can be carried out in an aqueous solution under conditions favoring endonuclease activity. Typically the solution is buffered to a pH of about 6.5 to 8.0. Mild temperatures, preferably about 20°C to about 45°C, more preferably physiological temperatures (25° to 40°C), are employed. Restriction 20 endonucleases normally require magnesium ions and, in some instances, cofactors (ATP and S-adenosyl methionine) or other agents for their activity. Therefore, a source of such ions, for instance inorganic 25 magnesium salts, and other agents, when required, are present in the digestion mixture. Suitable conditions are described by the manufacturer of the endonuclease and generally vary as to whether the endonuclease requires high, medium or low salt conditions for optimal activity.

30 The amount of DNA in the digestion mixture is typically in the range of 1% to 20% by weight. In most instances 5 to 20 µg of total DNA digested to completion provides an adequate sample for production of RFLP fragments. Excess endonuclease, preferably one to five 35 units/µg DNA, is used.

The set of fragments in the digest is preferably further processed to produce RFLP patterns which are analyzed. If desired, the digest can be purified by precipitation and resuspension as described by Kan et al, PNAS 75:5631-5635 (1978), prior to additional processing. That article is incorporated herein by reference in its entirety.

Once produced, the fragments are analyzed by well known methods. Preferably, the fragments are analyzed using electrophoresis. Gel electrophoresis methods are described in detail hereinafter. Capillary electrophoresis methods can be automated (as by using Model 207A analytical capillary electrophoresis system from Applied Biosystems of Foster City, CA) and are described in Chin et al, *American Biotechnology Laboratory News Edition*, December, 1989.

#### *Electrophoretic Separation of DNA Fragments*

Electrophoresis is the separation of DNA sequence fragments contained in a supporting medium by size and charge under the influence of an applied electric field. Gel sheets or slabs, e.g. agarose, agarose-acrylamide or polyacrylamide, are typically used for nucleotide sizing gels. The electrophoresis conditions affect the desired degree of resolution of the fragments. A degree of resolution that separates fragments that differ in size from one another by as little as 10 nucleotides is usually sufficient. Preferably, the gels will be capable of resolving fragments which differ by 3 to 5 nucleotides. However, for some purposes (where the differences in sequence length are large), discrimination of sequence differences of at least 100 nt may be sufficiently sensitive for the analysis.

Preparation and staining of analytical gels is well known. For example, a 3% Nusieve 1% agarose gel which is stained using ethidium bromide is described in

Boerwinkle et al, *PNAS*, 86:212-216 (1989). Detection of DNA in polyacrylamide gels using silver stain is described in Goldman et al, *Electrophoresis*, 3:24-26 (1982); Marshall, *Electrophoresis*, 4:269-272 (1983);  
5 Tegelstrom, *Electrophoresis*, 7:226-229 (1987); and Allen et al, *BioTechniques* 7:736-744 (1989). The method described by Allen et al, using large-pore size ultrathin-layer, rehydratable polyacrylamide gels stained with silver is preferred. Each of those  
10 articles is incorporated herein by reference in its entirety.

Size markers can be run on the same gel to permit estimation of the size of the restriction fragments. Comparison to one or more control sample(s) can be made in addition to or in place of the use of size markers.  
15 The size markers or control samples are usually run in one or both the lanes at the edge of the gel, and preferably, also in at least one central lane. In carrying out the electrophoresis, the DNA fragments are loaded onto one end of the gel slab (commonly called the "origin") and the fragments separate by electrically facilitated transport through the gel, with the shortest fragment electrophoresing from the origin towards the other (anode) end of the slab at the fastest rate. An agarose slab gel is typically electrophoresed using about 100 volts for 30 to 45 minutes. A polyacrylamide slab gel is typically electrophoresed using about 200 to 20  
20 25 1,200 volts for 45 to 60 minutes.

After electrophoresis, the gel is readied for visualization. The DNA fragments can be visualized by staining the gel with a nucleic acid-specific stain such as ethidium bromide or, preferably, with silver stain, which is not specific for DNA. Ethidium bromide staining is described in Boerwinkle et al, *supra*.  
30 35 Silver staining is described in Goldman et al, *supra*,

Marshall, *supra*, Tegelstrom, *supra*, and Allen et al., *supra*.

*Probes*

5        Allele-specific oligonucleotides or probes are used to identify DNA sequences which have regions that hybridize with the probe sequence. The amplified DNA sequences defined by a locus-specific primer pair can be used as probes in RFLP analyses using genomic DNA. U.S.  
10      Patent No. 4,582,788 (to Erlich, issued April 15, 1986) describes an exemplary HLA typing method based on analysis of RFLP patterns produced by genomic DNA. The analysis uses cDNA probes to analyze separated DNA fragments in a Southern blot type of analysis. As  
15      stated in the patent "[C]omplementary DNA probes that are specific to one (locus-specific) or more (multilocus) particular HLA DNA sequences involved in the polymorphism are essential components of the hybridization step of the typing method" (col. 6, l. 3-7).

20      The amplified DNA sequences of the present method can be used as probes in the method described in that patent or in the present method to detect the presence of an amplified DNA sequence of a particular allele. More specifically, an amplified DNA sequence having a known allele can be produced and used as a probe to detect the presence of the allele in sample DNA which is amplified by the present method.

25      Preferably, however, when a probe is used to distinguish alleles in the amplified DNA sequences of the present invention, the probe has a relatively short sequence (in comparison to the length of the amplified DNA sequence) which minimizes the sequence homology of other alleles of the locus with the probe sequence. That is, the probes will correspond to a region of the amplified DNA sequence which has the largest number of

nucleotide differences from the amplified DNA sequences of other alleles produced using that primer pair.

The probes can be labelled with a detectable atom, radical or ligand using known labeling techniques.

5 Radiolabels, usually  $^{32}\text{P}$ , are typically used. The probes can be labeled with  $^{32}\text{P}$  by nick translation with an  $\alpha$ - $^{32}\text{P}$ -dNTP (Rigby et al, *J. Mol. Biol.*, 113:237 (1977)) or other available procedures to make the locus-specific probes for use in the methods described in the 10 patent. The probes are preferably labeled with an enzyme, such as hydrogen peroxidase. Coupling enzyme labels to nucleotide sequences are well known. Each of the above references is incorporated herein by reference 15 in its entirety.

15 The analysis method known as "Southern blotting" that is described by Southern, *J. Mol. Biol.*, 98:503-517 (1975) is an analysis method that relies on the use of probes. In Southern blotting the DNA fragments are electrophoresed, transferred and affixed to a support 20 that binds nucleic acid, and hybridized with an appropriately labeled cDNA probe. Labeled hybrids are detected by autoradiography, or preferably, use of enzyme labels.

25 Reagents and conditions for blotting are described by Southern, supra; Wahl et al, *PNAS* 6:3683-3687 (1979); Kan et al, *PNAS*, supra, U.S. Pat. No. 4:302,204 and *Molecular Cloning: A Laboratory Manual* by Maniatis et al, Cold Spring Harbor Laboratory 1982. After the transfer is complete the paper is separated from the gel 30 and is dried. Hybridization (annealing) of the resolved single stranded DNA on the paper to an probe is effected by incubating the paper with the probe under hybridizing conditions. See Southern, supra; Kan et al, *PNAS*, supra and U.S. Pat. No. 4,302,204, col 5, line 8 et seq. 35 Complementary DNA probes specific for one allele, one locus (locus-specific) or more are essential components

of the hybridization step of the typing method. Locus-specific probes can be made by the amplification method for locus-specific amplified sequences, described above. The probes are made detectable by labeling as described  
5 above.

The final step in the Southern blotting method is identifying labeled hybrids on the paper (or gel in the solution hybridization embodiment). Autoradiography can be used to detect radiolabel-containing hybrids. Enzyme labels are detected by use of a color development system specific for the enzyme. In general, the enzyme cleaves a substrate, which cleavage either causes the substrate to develop or change color. The color can be visually perceptible in natural light or a fluorochrome which is excited by a known wavelength of light.  
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#### *Sequencing*

Genetic variations in amplified DNA sequences which reflect allelic difference in the sample DNA can also be detected by sequencing the amplified DNA sequences. Methods for sequencing oligonucleotide sequences are well known and are described in, for example, *Molecular Cloning: A Laboratory Manual* by Maniatis et al, Cold Spring Harbor Laboratory 1982. Currently, sequencing can be automated using a number  
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of commercially available instruments.

Due to the amount of time currently required to obtain sequencing information, other analysis methods, such as gel electrophoresis of the amplified DNA sequences or a restriction endonuclease digest thereof are preferred for clinical analyses.

#### *Kits*

As stated previously, the kits of this invention comprise one or more of the reagents used in the above described methods. In one embodiment, a kit comprises

at least one genetic locus-specific primer pair in a suitable container. Preferably the kit contains two or more locus-specific primer pairs. In one embodiment, the primer pairs are for different loci and are in separate containers. In another embodiment, the primer pairs are specific for the same locus. In that embodiment, the primer pairs will preferably be in the same container when specific for different alleles of the same genetic locus and in different containers when specific for different portions of the same allele sequence. Sets of primer pairs which are used sequentially can be provided in separate containers in one kit. The primers of each pair can be in separate containers, particularly when one primer is used in each set of primer pairs. However, each pair is preferably provided at a concentration which facilitates use of the primers at the concentrations required for all amplifications in which it will be used.

The primers can be provided in a small volume (e.g. 100  $\mu$ l) of a suitable solution such as sterile water or Tris buffer and can be frozen. Alternatively, the primers can be air dried.

In another embodiment, a kit comprises, in separate containers, two or more endonucleases useful in the methods of this invention. The kit will preferably contain a locus-specific combination of endonucleases. The endonucleases can be provided in a suitable solution such as normal saline or physiologic buffer with 50% glycerol (at about -20°C) to maintain enzymatic activity.

The kit can contain one or more locus-specific primer pairs together with locus-specific combinations of endonucleases and may additionally include a control. The control can be an amplified DNA sequence defined by a locus-specific primer pair or DNA having a known HLA type for a locus of interest.

Additional reagents such as amplification buffer, digestion buffer, a DNA polymerase and nucleotide triphosphates can be provided separately or in the kit. The kit may additionally contain gel preparation and staining reagents or preformed gels.

Analyses of exemplary genetic loci are described below.

Analysis of HLA Type

The present method of analysis of genetic variation in an amplified DNA sequence to determine allelic difference in sample DNA can be used to determine HLA type. Primer pairs that specifically amplify genomic DNA associated with one HLA locus are described in detail hereinafter. In a preferred embodiment, the primers define a DNA sequence that contains all exons that encode allelic variability associated with the HLA locus together with at least a portion of one of the adjacent intron sequences. For Class I loci, the variable exons are the second and third exons. For Class II loci, the variable exon is the second exon. The primers are preferably located so that a substantial portion of the amplified sequence corresponds to intron sequences.

The intron sequences provide restriction sites that, in comparison to cDNA sequences, provide additional information about the individual; e.g., the haplotype. Inclusion of exons within the amplified DNA sequences does not provide as many genetic variations that enable distinction between alleles as an intron sequence of the same length, particularly for constant exons. This additional intron sequence information is particularly valuable in paternity determinations and in forensic applications. It is also valuable in typing for transplant matching in that the variable lengths of

intron sequences included in the amplified sequence produced by the primers enables a distinction to be made between certain heterozygotes (two different alleles) and homozygotes (two copies of one allele).

5            Allelic differences in the DNA sequences of HLA loci are illustrated below. The tables illustrate the sequence homology of various alleles and indicate exemplary primer binding sites. Table 1 is an illustration of the alignment of the nucleotides of the Class I A2, A3, Ax, A24 (formerly referred to as A9), B27, B58 (formerly referred to as B17), C1, C2 and C3 allele sequences in intervening sequence (IVS) I and III. (The gene sequences and their numbering that are used in the tables and throughout the specification can be found in the Genbank and/or European Molecular Biology Laboratories (EMBL) sequence databanks. Those sequences are incorporated herein by reference in their entirety.) Underlined nucleotides represent the regions of the sequence to which exemplary locus-specific or Class I-specific primers bind.

10           Table 2 illustrates the alignment of the nucleotides in IVS I and II of the DQA3 (now DQA1 0301), DQA1.2 (now DQA1 0102) and DQA4.1 (now DQA1 0501) alleles of the DQA1 locus (formerly referred to as the DR4, DR6 and DR3 alleles of the DQA1 locus, respectively). Underlined nucleotides represent the regions of the sequence to which exemplary DQA1 locus-specific primers bind.

15           Table 3 illustrates the alignment of the nucleotides in IVS I, exon 2 and IVS II of two individuals having the DQw1<sub>v</sub> allele (designated hereinafter as DQw1<sub>v</sub>a and DQw1<sub>v</sub>b for the upper and lower sequences in the table, respectively), the DQw2 and DQw8 alleles of the DQB1 locus. Nucleotides indicated in the DQw1<sub>v</sub>b, DQw2 and DQw8 allele sequences are those which differ from the DQw1<sub>v</sub>a sequence. Exon 2 begins and ends

at nt 599 and nt 870 of the DQw1<sub>y</sub>a allele sequence, respectively. Underlined nucleotides represent the regions of the sequence to which exemplary DQB1 locus-specific primers bind.

5           Table 4 illustrates the alignment of the nucleotides in IVS I, exon 2 and IVS II of the DPB4.1, DPB9, New and DPW3 alleles of the DPB1 locus. Nucleotides indicated in the DPB9, New and DPW3 allele sequences are those which differ from the DPB4.1 10 sequence. Exon 2 begins and ends at nt 7644 and nt 7907 of the DPB4.1 allele sequence, respectively. Underlined nucleotides represent the regions of the sequence to which exemplary DPB1 locus-specific primers bind.

TABLE 1

## Class I Seq

	C1	1	GATTACCAATATTGTGCGACCTACTGTATCAATAAAC	
5	C2	1		T
	C1	38	AAAAAGGAAACTGGTCTCTATGAGAACTCTACCTGGTGCTTCAGACAA	
	C2	38		G G
	C1	88	CACTCACCAAGTTAAAGAGAAAACCTCCTGACTCTACACGTCCATTCCC	
	C2	88		
10	B27	1	GAGCTCACTCTCTGGCATCAAGTTIC	TOCGTG
	C1	138	AGGGCGAGCTCACTGTCTGGCAGCAAGTTCCCCATGGTCGAGTTTCCTG	
	C2	138		T
	A2	1	AAGCTTACTCTCTGGCACCAAAC	TOCATGGGATGATTTTCCCTTCC TAG
	B27	32		ATCAGTTTCCCT
	C1	188	TACAAGAGTCCAAGGGAGAGGTAAAGTGTCTTT	AT
	C2	187		TTTGCTGGATGTAG
15	A2	50	AAGAGTCCAGGTGGACAGTAA	GGAGTGGAGT CAGGGAGTC
	B27	44	ACACAAGA TCCAAGAGGAGAGTAA	GGAGT GAG AGGCAGGGAGTC
	C1	238	TTTAATATTACCT GAGGTAAAGGTAA	GGC AAAGAGTGGG AGGCAGGGAGTC
	C2	237		C - G
20	A2	98	CAGTTCCAGGGACAGAGATTACGGGATAAAAAGTGAAGGAGAGGGACG	GGGOCAT
	B27	91	CAGTT CAGGGACAGGGATTCCAGGAGAGAAGTGAAGGGGAAGC	GGG TGGGC
	C1	288	CAGTT CAGGGACAGGGATTCCAGGAGAGAAG	TGAAGGGGAAG GGGCTGGGCG
	C2	288		
25	A2	149	GGCGAG GGTTCCTCCCTTCT CAGACAGCTC TTGGGCGA A GAC	
	B27	141	GGCACTGGGGGTCTCTCCTGGTTCCACAGACAGATCCTTGTGCC	GGAC
	C1	338	CAGCC TGGGGTCTCTCCTGGTTCCACAGACAGATCCTTG	GCC AGGAC
	C2	337		-- GG
	A2	195	TCAGGGAGACATTGAGACAGAGC	GCTGGCACAGAACAGAGGGTCAGGG
	B27	191	TCAGGCAGACAGTGTGACAAGAGGT	GGTGTAGGAGAAGAGGGATCAGG
	C1	388	TCAGGCACACAGTGTGACAAGATGCTTGTGTAGGAGAAGAGGGATCAG	
	C2	387		G
30	A2	246	CGAA GTCCAGGGCCCCAGCGTTGGCTCTCAGGGTCTCAGGCCCGAAGG	
	A3	1		
	Ax	1		
	A24	1		
	B27	241	ACGAACGTCCAAGGCCCCGGCG	CGG TCTCAGGGTCTCAGGGCTCCGAGAG
	C1	438	ACGAA GTGCCAGGTCCCCGGCG	GGGTTCTCAGGGTCTCAGGGCTCCAAGGG
	C2	438		-A

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3

	A2	296	CGGTGTATGGATTGGGAGTCGCCAGCCTTGGGATTCCCCAACTCCGC	AGTT	
	A3	9	T	A	
	Ax	9		TG	G C
5	A24	11			T
	B27	291	CCTTGTCTGCATTGGGAGGGCGCACAGTTGGG	TTCCCCACTCCACGAGTT	
	C1	488	<u>CGTGTCTGCACTGGGAGGCCGCCGTTGAGGATTCCTCACTCCCC</u> TGA		
	C2	488			
	A2	348	TCTTTCTCCC	TCTCCAACCTATGTAGGGTCTTCTTCTGGAT	ACTCAC
10	A3	60	CTG	C	A G
	Ax	61	C	— A	GC AC C
	A24	61		TG-	—
	B27	344	TCACTTCT	TCTCCAACCTATGTCGGTCTTCTTCCAGGAT	ACTCGT
	C1	538	G TTCACCTCTCTCCAACCTGCGTCGGTCTTCTTGAAT	ACTCAT	
	C2	538	T	A	
	C3	1		T G	G
15	A2	399	GACCGGGACCCAGTTCTCACTCCATTGGGTGTCGGTTCC	AGAGAAG C	
	A3	114			
	Ax	109	A A	T C A	— T
	A24	111			G
	B27	392	GACCGGTCCCCATTTC	CACTCCATTGGGTGTCGGGT	GTCTAGAGAAG C
	B58	1			
	C1	588	GACCGGTCCCCAATTCCCACTCCATTGGGTGTCGGGT	TCT AGAAG C	
	C2	589			AG
20	C3	36		—ACNN	G
	A2	449	CAATCAGTGTCTCGCGGTGCGGGTCTAAAGT	CCGCACG	
	A3	164		T C	
	Ax	159	G C C	C	C
	A24	161	A	T	
25	B27	442	CAATCAGTGTCTCGGGTCCCCAGTTCTAAAGT	CCCCACG	
	B58	12			
	C1	635	CAATCAGCGTCTCGCACTCCATTGGGTCTAAAGTCCC	CAGT	
	C2	637	C		
	C3	87	GG	G	
	A2	489	<u>CACCCACCGGACTCAGA TTCTCCCCAGACCGGAGGATGGC</u>		C
	A3	204		TCGTGGAGACCAGGC	
	Ax	199		T	G
30	A24	201			
	B27	482	CACCCACCGGACTCAGA ATCTCTCAGACCGGAG	ATGCG	G
	B58	52			
	C1	675	CACCCACCGGACTCAGA TTCTCCCCAGACCGGAG	ATGCG	G
	C2	677	G		
	C3	127			

1st EXON

	A2	532	GTCATGGCGCCCGAACCCTCGTCTGCTACTCTCGGGGGCTC		
	A3	262		C	C
5	Ax	242	C	C	
	A24	244	G	A	C
	B27	524	<u>GTCACGGCGCCCGAACCCTCGTCTGCTCTGGGGGCAG</u>		
	B58	94		G	
	C1	717	GTCATGGCGCCCGAACCCTCATCGTCTGCTCTGGGAGCCC		
	C2	719			
	C3	169		G	
10	A2	574	<u>TGGCCCTGACCCAGACCTGGCGG</u>		
	A3	305			
	Ax	285		C	
	A24	287		A	
	B27	567	TGGCCCTGACCGAGACCTGGCGT		
	B58	137		C	
	C1	760	TGGCCCTGACCGAGACCTGGCGT		
15	C2	762			
	C3	212		G	
			IVS1		
	A2	599	GTGAGTGCAGGGTCAGGG AGGGAAACGCC TCTGT GGGGAGAACGCAACGGGCC G		
	A3	329		C AC	C G T
	Ax	309	A T C	T-G — — —	G NG G CG
	A24	311		TCG C C	G CG
20	B27	591	GTGAGTGCAGGGTCAGGGAGGGAAATG GCC TCTGT GGGGAGGAGCGAGGGGA CG		
	B58	161		G —	C
	C1	784	GTGAGTGCAGGGTCAGGG AGGGAAACGCC TCT GTGGAGAGGAACGAGGTGCCCG		
	C2	786			G G
	C3	236		T T	G G
25	A2	652	CCTGGC GGGGGCGCAGGACCGGGAGGCCGCCGGAGGGAGGGTCGGGGGGCTCAG		
	A3	383		G G	C
	Ax	357	C G T	A G A	
	A24	367		A	
	B27	645	CAGGC GGGGGCGCAGGACCGGGAGGCCGCCGGAGGGAGGGTCGGGGGGCTCAG		
	B58	215		T A	
	C1	838	CCCGGC AGG CGCAGGACCGGGAGGCCGCCAGGGAGGGAGGGTCGGGGGGCTCAG		
	C2	840		G G — AGC	
	C3	291		GGA G	
30	A2	711	CCACTCTCGTCCCCAG		
	A3	442	— G — C		
	Ax	417	TC CT		
	A24	426			
	B27	703	CCACTCTCGCAGG		
	B58	273			
35	C1	895	CCACTCTCGCAGG		
	C2	898		T	
	C3	351		—	

## IVS3

	A2	1515	GTACCAGGGGCCACGGGGCGCTCCCTGATCGCTGTAGATCTCCGGCTGGCTCCC				
	A3	1245		-			
	Ax	1222	C ACA	-			
5	A24	1228			G		
	B27	1508	GTACCAGGGCAGTGGGGAGCCTCCCATCTCTATAGTCGCOGGGATGGCTCCC				
	B58	1082					
	C1	1704	GTACCAGGGCAGTGGGGAGCCTCCCATCTCCCGTAGATCTCCGGCATGGCTCCC				
	C2	1705		T	G		
	C3	1155		T	G		
10	A2	1574	ACAAGGAGGGAGACAATTGGGACCAACACTAGAATATGCCCTCCCTCTGGT				
	A3	1303		C C G A T T			
	Ax	1280	A A A	T			
	A24	1287	C				
	B27	1567	ACGAGAACAGGAGGAAATGGGATCAGCGCTAGAATGTCGCCCTCCCTGAAT				
	B58	1141					
	C1	1763	ACGAGGAGGGAGGAAATGGGATCAGCGCTAGAATATGCCCTCCCTGAAAT				
	C2	1764					
15	C3	1213					
	A2	1627	CCTGAGGGAGAGGAATCTCTGGTTCCAGATCTGTACCAAGAGGTGA				
	A3	1356	T T T	- GA G			
	Ax	1333	T	T			
	A24	1341	T				
	B27	1620	GGAGAACATGGCATGAGTTTCTGAGITTC				
	B58	1194					
	C1	1816	GGAGAACATGGCATGAGTTTCTGAGITTC				
	C2	1817					
	C3	1266					
	A2	1678	CTCTGAGGTCCGCCCTGCTCTGA CACAATTAAAGGGATAAAATCTCTGAAGGA				
	A3	1406	T G A A -G				
	Ax	1372	G -		G G -		
20	A24	1392				C	
	B27	1649	CTCTGAGGGCCCOCTCTCTCT AGGACAATTAAAGGGATGACGTCTCTGAGGAA				
	B58	1223					
	C1	1845	CTCTGAGGGCCCOCTCTGCTCT AGGACAATTAAAGGGATGAAGTCTTGAGGAA				
	C2	1846					
	C3	1295		G		A	
25	A2	1733	ATGACGGG AAGACGATCCCTCGAATACTGATGAGTGGTTCCCTTGTACAC				
	A3	1460	G T T G T G			G	
	Ax	1426	ATGAA G A G				
	A24	1447	A		C		
	B27	1704	ATGGAGGGAAAGACAGTCCTAGAATACTGATCAGGGTCCCTTGTACCC				
	B58	1278					
	C1	1900	ATGGAGGGAAAGACAGTCCTGGAATACTGATCAGGGTCCCTTGTACCA				
	C2	1901					
30	C3	1351		A			

	A2	1783	ACACAGGCAGCAGCCTTGGG CCCG	TGACTTTCTCTCAGGCCTTGTCTCTGC
	A3	1510	—C GA G	
	Ax	1477	—T	C
	A24	1497	—C	A
5	B27	1755	CTGCAGCAGCCTTGGGAACCG	TGACTTTCTCTCAGGCCTTGTACAGC
	B58	1329		T T
	C1	1951	CTTGACCCTGCAGCAGCTGTGGTCAGGCTGCTGACCTTT	CTCTCAGGCCTTGTCTCTGC
	C2	1952		
	C3	1411	—	
	A2	1837	TTCACACTCAATGTGTGTGGGGTCTGAGTCCAGCACCTCTGAGTCCTTCAGCC	
10	A3	1560		C
	Ax	1528	C	—
	A24	1547		C
	B27	1806	CTCACACTCAGTGTGTTGGGCTCTGATTCCAGCACCTCTGAGTCACITTTACC	
	B58	1380		
	C1	2013	CTCACGTTCAATGTGTTGAAGGTTGATTAGCTTCTGAGTCCTTCGGCC	
	C2	2014		
15	C3	1464	C	
	A2	1891	TCCACTCAGGTCAAGGACCAGAAGTCGCTGTCCCTCTCAGGGACTAGAA	TTTCCACCGAATAG
	A3	1614		TC A —
	Ax	1567		T
	A24	1600		A —
	B27	1860	TCCACTCAGATCAGGAGCAGAAGTCCTGTCCCTCAGAGACT	CGAACCTTCCAATGAATAG
	B58	1434		
20	C1	2067	TCCACTCAGGTCAAGGACCAGAAGTCGCTGTCCCTCAGAGACTAGAACTTCCAATGAATAG	
	C2	2068		
	C3	1518		
	A2	1955	GAGATTATCCCAGGTGCCTGTGTCCAGGCTGGTGTCTGGGTTCTGTGCTCCCTCCCCA	
	A3	1664	—	
	Ax	1632	T T C T T	
	A24	1650	— A	A T G
25	B27	1925	GAGATTATCCCAGGTGCCTGGTGCAGGCTGGTGTCTGGGTTCTGTGCCAC CTCCCCA	
	B58	1499		
	C1	2132	GAGATTATCCCAGGTGCCTGTGTCCAGGCTGGCGTCTGGGTTCTGTGCCCTTCCCCA	
	C2	2133		
	C3	1583		

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A2	2014	TCCCAGGTGTOCTGTOCATTCTCAAGA	TAGGCCACATGTGTGCTGGAGGAGTGTCCCATG		
A3	1721	G	G	C T	
Ax	1691	C T CA	A	G C T	
A24	1706			G CA T	
5	B27	1983 CCCCAGGTGTOCTGTOCATTCTC	<u>AGGCTGGTACATGGGTGGTCCTAGGGTGTCCCATG</u>		
B58	1557	A			
C1	2191	CCCCAGGTGTOCTGTOCATTCTC	<u>AGGATGGTACATGGGGCTGTTGGAGTGTGCGAAG</u>		
C2	2192		A		
C3	1642		G		
10	A2	2073 ACAGATCGAAAATGCCTGAATGATCTGACTCT	TOCTGACAG	2113	
A3	1780	GC	TT	C T	1820
Ax	1750	GC	TT	TT C T	1791
A24	1765	G GAAAA	-	C T	1784
B27	2042 AGAGATGCAAAGCGCCTGAATTCTGACTCTCCAT	CAG	2083		
B58	1616			1656	
C1	2250 AGAGATAACAAAGTGCTGAATTCTGACTCTCCGT	CAG	2290		
C2	2251		G	2292	
C3	1701			1741	

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TABLE 2

DQAI Seq

	A3	1	GATCTCTGTAGAATGTC <u>CTGTTCTGAGCCAGTCCTGAGAGGAAGTATAATCAA</u>											
5	A1.2	1		G	A									
	A4.1	1	C	G									A	A
													C	G
	A3	61	TTTGTTATTAA <u>CTGATGAAAGAATTAAAGTGAAGATAAACCTTAGGAAGC AGAGGAAAGT</u>											
	A1.2	61		CA									T	C
	A4.1	61											G	T
													C	A
10	A3	121	TAA	TCTATGACTAAGAAAGTTAAGTACTCTGATAACTCATTCA <u>TCATTCCCTTCT</u>										
	A1.2	122	A CCTAA	T C		C	A	A						
	A4.1	122	A CCTAA	C		C	A	CA A						
	A3	172	TTTGTTCATTACATT	ATTTAAC <u>TAACAAAGTCTATGATGTGCCAGGCCTCTCAGGAAATA</u>										
	A1.2	178		A		T	C	C					A	
	A4.1	178		A	G		T	CG					A	
15	A3	230	GTGAAAATTGG	CACGCGATATTCTGCC <u>CTTGTGTAGCACACACCCGTAGTGGGAAAG</u>										
	A1.2	236	A	A T									G	TAG
	A4.1	237	A	C	A	TT T							G	TTA
	A3	286	AA GTGCACTTTAACC	GGACA <u>ACTATCAACACGAAGCGGGAGGAAGCAGGGG</u>										
	A1.2	293	A		T		C	T	A					
	A4.1	294	A C	A			C	AT	A T					
20	A3	339	CTGGAAATGTCCACAGA	CTTGC <u>AAA GACAAAGCCCATAATATCTGAAAGTCAG</u>										
	A1.2	347		G	AA	TG							T	
	A4.1	348	T	G	G		TG		G			T		
	A3	394	TTTCTTC	CATCATTTGTGTATTAAGGTT <u>CTTTATTCCCTGTTCTCTGCCTTCCT</u>										
	A1.2	403	G CT			C	T						C	
	A4.1	403	CT	TCAT			G C						CA	
25	A3	450	<u>GCTTGTCA</u> TCTCACTCATCAGCTGACC <u>ATGTTGCCTCTACGGTGTAAAC</u> TTGTACCAG											
	A1.2	459			C								GT	
	A4.1	462			C	C							T	
	A3	510	TCTTATGGCC	CT <u>CTGGCAGTACAGCCATGAATTGATGGAGACGAGGAGTTCTAT</u>										
	A1.2	519	T	C		C	C						T	C
	A4.1	522	C		C	C							T	C
														C
	A3	567	GTGGACCTGGAGAGGA	AGGA <u>ACTGTCTGGCAGTTGCCTCTGTTCCGCAGATTAA</u>										
	A1.2	576		C	G	G	GA	A	A				G	
	A4.1	579		G			TGT		G TC	A ACA				
30	A3	622	GAAGATTGACCCG	CAATTG <u>ACTGACAAACATCGCTGTGCTAAAACATAACTTGA</u>										
	A1.2	631	G T	GGG		G	G						GC	C
	A4.1	634	---										C	
	A3	679	ACATCGTGA	TTAA <u>ACGCTCCA</u> ACT <u>CTACCGCTGCTACCA</u> ATGGTATGTGTCCACCATTCTG										
	A1.2	688	A		A								C	
	A4.1	688	GTC										A	A

DQA1 Seq (cont.)

5	A3      740	CCTTTCTTAC	TGATTATCCCTTATACCAAGTTCATTTCTTT	
	A1.2    749	C            TTAA A GC	CC            G	C
	A4.1    749	CC	C	A

A3 789 CCAAGAGGTCCCCAGATC 806  
A1.2 802 819  
A4.1 798 815

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169,0018

TABLE 3

DQB1 Seq

	1	AAGCTTGTGCTCTTCATGAATAAATGTCTCATCTAGGACTCAGAGGT						
5			GG	T	T		A	
						G		
	51	GTAGG TCCTTCCAACATAGAAGGGAGTGA	TT			ACCTCAACGGG ACTTGGGA G		
		C AC C TTT TA C CA AC	A T			TT CA C AT C		
10	101	GGTAAATCTAGGCATGGGAACGAAGGTATTACCCAGGGACCAAGAGAA						A
			C			G		
15	151	TACCGTGTCAAGAACGAGGCCAGGCTTAATTCTGGACCTATCTCGTCAT						
		G A G - A T	A A	T	T	G CG	A	
	201	TCGGTGAACCTCAGATTATGTGGATAACTTATCTCTGAGGTATCCA						
		C G G	C A G		T		C T	
20	251	GGAGCTCATGAAAATGGGATTTCATGCGAGAACGCCCTGAT CCCTCTA						
		C G A	CA G			G	T	
25	301	AGTGCAGAGGTGCATGTAAAATCAGCCGACTGCCTCTCGCTGGGTICA						
		C	CT			A T		
	351	CAGGCTCAGGCAGGGACAGGGCTTTCCTCCCTTCCTGGATGTAGGAAGG						
		CG A	C			CC		
30					G	CC C		
	401	C AGATTCAGAAGGCCGCAAAGAAGGCGGGAGAGCTGGCAGAGCCGCC						
		CG C A C CG G	C G G			G G	- N N N	
	451	GGGAGGATCCCAGGTCTGGAGGCCAGGCACGGGCGGGCGGGAACTGGAG						
35			C A A				T T	

501 GTCGCCGGGCGGTCCACAGCTCCAGGCGGGTCAGGGCGGCTGG  
 5 T G T  
 551 GGGCGGGCGGCGTGGGCC TGACTGACCGGCCGGTGATCCCCGCAGAG  
 10 A - GCA --  
 601 GATTTCGTGTACCAGTTAAGGGCATGTGCTACTTCACCAACGGACGGA  
 15 A  
 651 GCGCGTGCCTTGTAAACCAGACACATCTATAACCGAGAGGGAGTACCGC  
 20 G G AG A AT T  
 701 GCTTCGACAGCGACGTGGGGGTGTACCGGGCGTGACGCCAGGGCGG  
 25 A T T T T C  
 751 CCTGTTGCCGAGTACTGGAACAGGCCAGAAGGAAGTCTGGAGAGGACCG  
 30 CC CA AA  
 801 GGCAGAGTTGGA CACGGTGTGCAGACACAACTACGAGGTGGGTACCGC  
 25 C G G C T A C T A  
 851 GGATCCTGCAGAGGAGAGGTGAGCTTCGTCGCCCTCGTGAGGCC ACCC  
 35 C C T C C G G - T T C G C C  
 C C T C C G G G - T T C G C C T  
 901 TTGGCCGGACCOGAGTCTCTGTGCCGGAGGGCG ATGGGGCGAGGTC  
 30 — A C A G CAA G CCG T T C  
 A G A C G CAA G CCG T T C  
 951 TCTGAAATCTGAGGCCAGTCATTCCACCCCAGGGAAAGGAGGCGGCGG  
 35 — C — C TT CGG — CTG C — A A

1001 CGGGGGTGGTGGGGGCAGGTGCATCGGAGGGGGGGACCTAGGGCAGAG  
CGGT - C T A  
5 1051 CAGGGGGACAAGCAGAGTTGCCAGGCTGCCTAGTGTCCCCCCCAGCCTC  
G T A T G - T  
1101 CTCTCGTCGTCGGCCTCGTCCCTCTGCTCTGGACGTTCTCGCCTCGTGCCT  
C C C - T  
10 1151 TATGGGTTTGCTTCTCGTGCCTAACCTTCGCTAAGCAGTTCTCTCTGCC  
TA  
1201 CCCAGTCCCCACCCCTCTTCCCCCTGCCCGCCGGCCTCGCTAGCACTGCC  
A TT G C CG G  
15 1251 ACCCAGCAAGGCCACAGTCGCGCATTCGGCGCA GGAAGCTT 1292  
T CG  
G T CTA A AGC CATG AGTGGGAAGCTT  
20  
25  
30  
35

TABLE 4

DPB1 Seq

DPB4.1 7546

GGGAAGATTGGGAAGAAATCGTTAATAT

5 DPB4.1 7574 TGAGAGAGAGAGGGAGAAAGAGGATTAGATGAGAGTGGCGCTCCGCTCATGTCGCC

DPB4.1 7634 CTOCCCGCAGAGAATTACCTTTCAGGACGGCAGGAATGCTACCGTTAACGGACA

DPB9           GGAT                   G GCA           TT

New           GGAT                   G GCA           TT

DPw3

10

BPB4.1 7694 CAGCGCTTCCTCGAGAGATACTACAAACCGGAGGAGTTCGCGCGCTCGACAGCGAC

DPB9

T

New

T

DPw3

15

DPB4.1 7754 GTGGGGAGTTCCGGCGGTGACGGAGCTGGGCGGACTGCTGCAGAGTACTGGAACAGC

DPB9

A A C

New

A A C

DPw3

20

DPB4.1 7814 CAGAAGGACATCCTGGAGGAGAACGGGCAGTGCGGACAGGATGTGCAGACACAAC

DPB9

G

G A

New

G A

DPw3

C

G A

25

DPB4.1 7874 GAGCTGGGGGGCCOCATGACOCTGCAGGGGGAGGTGAGTGAGGGCTTGGGCCGGT

DPB9

A A G G

New

A A G G

DPw3

A A G G

30

DPB4.1 7934 CCAGGGCAGCCCCGGGGCCGTGCCAG

35

## Primers for HLA loci

Exemplary HLA locus-specific primers are listed below. Each of the primers hybridizes with at least about 15 consecutive nucleotides of the designated region of the allele sequence. The designation of an exemplary preferred primer together with its sequence is also shown. For many of the primers, the sequence is not identical for all of the other alleles of the locus. For each of the following preferred primers, additional preferred primers have sequences which correspond to the sequences of the homologous region of other alleles of the locus or to their complements.

In one embodiment, Class I loci are amplified by using an A, B or C locus-specific primer together with a Class I locus-specific primer. The Class I primer preferably hybridizes with IVS III sequences (or their complements) or, more preferably, with IVS I sequences (or their complements). The term "Class I-specific primer", as used herein, means that the primer hybridizes with an allele sequence (or its complement) for at least two different Class I loci and does not hybridize with Class II locus allele sequences under the conditions used. Preferably, the Class I primer hybridizes with at least one allele of each of the A, B and C loci. More preferably, the Class I primer hybridizes with a plurality of, most preferably all of, the Class I allele loci or their complements. Exemplary Class I locus-specific primers are also listed below.

## HLA Primers

### A locus-specific primers

allelic location: nt 1735-1757 of A3

designation: SGD009.AIVS3.R2NP

sequence: CATGTGGCCATCTTGAGAATGGA

allelic location: nt 1541-1564 of A2  
designation: SGD006.AIVS3.R1NP  
sequence: GCCCGGGAGATCTACAGGCGATCA

5 allelic location: nt 1533-1553 of A2  
designation: A2.1  
sequence: CGCCTCCCTGATGCCCTGTAG

10 allelic location: nt 1667-1685 of A2  
designation: A2.2  
sequence: CCAGAGAGTGACTCTGAGG

15 allelic location: nt 1704-1717 of A2  
designation: A2.3  
sequence: CACAATTAAGGGAT

B locus-specific primers

20 allelic location: nt 1108-1131 of B17  
designation: SGD007.BIVS3.R1NP  
sequence: TCCCCGGCGACCTATAAGGAGATGG

25 allelic location: nt 1582-1604 of B17  
designation: SGD010.BIVS3.R2NP  
sequence: CTAGGACCACCCATGTGACCAGC

allelic location: nt 500-528 of B27  
designation: B2.1  
sequence: ATCTCCTCAGACGCCGAGATGCGTCAC

30 allelic location: nt 545-566 of B27  
designation: B2.2  
sequence: CTCCTGCTGCTCTGGGGGGCAG

allelic location: nt 1852-1876 of B27  
designation: B2.3  
sequence: ACTTTACCTCCACTCAGATCAGGAG

5 allelic location: nt 1945-1976 of B27  
designation: B2.4  
sequence: CGTCCAGGCTGGTGTCTGGTTCTGTGCCCT

10 allelic location: nt 2009-2031 of B27  
designation: B2.5  
sequence: CTGGTCACATGGGTGGTCCTAGG

15 allelic location: nt 2054-2079 of B27  
designation: B2.6  
sequence: CGCCTGAATTCTGACTCTCCCAT

C locus-specific primers

20 allelic location: nt 1182-1204 of C3  
designation: SGD008.CIVS3.R1NP  
sequence: ATCCCAGGAGATCTACAGGAGATG

25 allelic location: nt 1665-1687 of C3  
designation: SGD011.CIVS3.R2NP  
sequence: AACAGCGCCATGTGACCATCCT

allelic location: nt 499-525 of C1  
designation: C2.1  
sequence: CTGGGGAGGCAGCCGCGTTGAGGATTCT

30 allelic location: nt 642-674 of C1  
designation: C2.2  
sequence: CGTCTCCGCAGTCCCGTTCTAAAGTTCCCAGT

allelic location: nt 738-755 of C1  
designation: C2.3  
sequence: ATCCTCGTGCCTCTCGGGA

5 allelic location: nt 1970-1987 of C1  
designation: C2.4  
sequence: TGTGGTCAGGCTGCTGAC

10 allelic location: nt 2032-2051 of C1  
designation: C2.5  
sequence: AAGGTTTGATTCCAGCTT

15 allelic location: nt 2180-2217 of C1  
designation: C2.6  
sequence: CCCCTTCCCCACCCCAGGTGTTCCATTCTTCAGGA

20 allelic location: nt 2222-2245 of C1  
designation: C2.7  
sequence: CACATGGGCGCTGTTGGAGTGTGCG

Class I loci-specific primers

25 allelic location: nt 599-620 of A2  
designation: SGD005.IIIVS1.LNP  
sequence: GTGAGTGCAGGGTCAGGGAGGGA

allelic location: nt 489-506 of A2  
designation: 1.1  
sequence: CACCCACCGGGACTCAGA

30 allelic location: nt 574-595 of A2  
designation: 1.2  
sequence: TGGCCCTGACCCAGACCTGGGC

allelic location: nt 691-711 of A2  
designation: 1.3  
sequence: GAGGGTCGGGCGGGTCTCAGC

5 allelic location: nt 1816-1831 of A2  
designation: 1.4  
sequence: CTCTCAGGCCTTGTTC

10 allelic location: nt 1980-1923 of A2  
designation: 1.5  
sequence: CAGAAGTCGCTGTTCC

DQA1 locus-specific primers

15 allelic location: nt 23-41 of DQA3  
designation: SGD001.DQA1.LNP  
sequence: TTCTGAGCCAGTCCTGAGA

20 allelic location: nt 45-64 of DQA3  
designation: DQA3 E1a  
sequence: TTGCCCTGACCACCGTGATG

25 allelic location: nt 444-463 of DQA3  
designation: DQA3 E1b  
sequence: CTTCCCTGCTTGTCACTTTCA

allelic location: nt 536-553 of DQA3  
designation: DQA3 E1c  
sequence: CCATGAATTGATGGAGA

30 allelic location: nt 705-723 of DQA3  
designation: DQA3 E1d  
sequence: ACCGCTGCTACCAATGGTA

allelic location: nt 789-806 of DQA3  
designation: SGD003.DQA1.RNP  
sequence: CCAAGAGGTCCCCAGATC

5       DRA locus-specific primers

allelic location: nt 49-68 of DRA HUMMHDRAM (1183 nt sequence, Accession No. K01171)  
designation: DRA E1  
sequence: TCATCATAGCTGTGCTGATG

10      allelic location: nt 98-118 of DRA HUMMHDRAM (1183 nt sequence, Accession No. K01171)  
designation: DRA 5'E2 (5' indicates the primer is used as the 5' primer)  
sequence: AGAACATGTGATCATCCAGGC

allelic location: nt 319-341 of DRA HUMMHDRAM (1183 nt sequence, Accession No. K01171)  
designation: DRA 3'E2  
sequence: CCAACTATACTCCGATCACCAAT

DRB locus-specific primers

allelic location: nt 79-101 of DRB HUMMHDRC (1153 nt sequence, Accession No. K01171)  
designation: DRB E1  
sequence: TGACAGTGACACTGATGGTGCTG

25      allelic location: nt 123-143 of DRB HUMMHDRC (1153 nt sequence, Accession No. K01171)  
designation: DRB 5'E2  
sequence: GGGGACACCCGACCACGTTTC

allelic location: nt 357-378 of DRB HUMMHDR (1153 nt sequence, Accession No. K01171)  
designation: DRB 3'E2  
sequence: TGCAGACACAACCTACGGGGTTG

5

DQB1 locus-specific primers

allelic location: nt 509-532 DQB1 DQw1<sub>v</sub>a  
designation: DQB E1  
sequence: TGGCTGAGGGCAGAGACTCTCCC

10

allelic location: nt 628-647 of DQB1 DQw1<sub>v</sub>a  
designation: DQB 5'E2  
sequence: TGCTACTTCACCAACGGGAC

15

allelic location: nt 816-834 of DQB1 DQw1<sub>v</sub>a  
designation: DQB 3'E2  
sequence: GGTGTGCACACACAAC

20

allelic location: nt 124-152 of DQB1 DQw1<sub>v</sub>a  
designation: DQB 5'IVS1a  
sequence: AGGTATTTACCCAGGGACCAAGAGAT

25

allelic location: nt 314-340 of DQB1 DQw1<sub>v</sub>a  
designation: DQB 5'IVS1b  
sequence: ATGTAAAATCAGCCCCACTGCCTCTTC

30

allelic location: nt 1140-1166 of DQB1 DQw1<sub>v</sub>a  
designation: DQB 3'IVS2  
sequence: GCCTCGTGCCTTATGCGTTGCCTCCT

35

DPB1 locus-specific primers

allelic location: nt 6116-6136 of DPB1 4.1  
designation: DPB E1  
sequence: TGAGGTTAATAAACTGGAGAA

allelic location: nt 7604-7624 of DPB1 4.1  
designation: DPB 5'IVS1  
sequence: GAGAGTGGCGCCTCCGCTCAT

5 allelic location: nt 7910-7929 of DPB1 4.1  
designation: DPB 3'IVS2  
sequence: GAGTGAGGGCTTTGGGCCGG

Primer pairs for HLA analyses

10 It is well understood that for each primer pair, the 5' upstream primer hybridizes with the 5' end of the sequence to be amplified and the 3' downstream primer hybridizes with the complement of the 3' end of the sequence. The primers amplify a sequence between the regions of the DNA to which the primers bind and its complementary sequence including the regions to which the primers bind. Therefore, for each of the primers described above, whether the primer binds to the HLA-encoding strand or its complement depends on whether the primer functions as the 5' upstream primer or the 3' downstream primer for that particular primer pair.

15  
20  
25 In one embodiment, a Class I locus-specific primer pair includes a Class I locus-specific primer and an A, B or C locus-specific primer. Preferably, the Class I locus-specific primer is the 5' upstream primer and hybridizes with a portion of the complement of IVS I. In that case, the locus-specific primer is preferably the 3' downstream primer and hybridizes with IVS III. The primer pairs amplify a sequence of about 1.0 to about 1.5 Kb.

30  
35 In another embodiment, the primer pair comprises two locus-specific primers that amplify a DNA sequence that does not include the variable exon(s). In one example of that embodiment, the 3' downstream primer and the 5' upstream primer are Class I locus-specific primers that hybridize with IVS III and its complement,

respectively. In that case a sequence of about 0.5 Kb corresponding to the intron sequence is amplified.

Preferably, locus-specific primers for the particular locus, rather than for the HLA class, are used for each primer of the primer pair. Due to differences in the Class II gene sequences, locus-specific primers which are specific for only one locus participate in amplifying the DRB, DQA1, DQB and DPB loci. Therefore, for each of the preferred Class II locus primer pairs, each primer of the pair participates in amplifying only the designated locus and no other Class II loci.

#### Analytical methods

In one embodiment, the amplified sequence includes sufficient intron sequences to encompass length polymorphisms. The primer-defined length polymorphisms (PDLPs) are indicative of the HLA locus allele in the sample. For some HLA loci, use of a single primer pair produces primer-defined length polymorphisms that distinguish between some of the alleles of the locus. For other loci, two or more pairs of primers are used in separate amplifications to distinguish the alleles. For other loci, the amplified DNA sequence is cleaved with one or more restriction endonucleases to distinguish the alleles. The primer-defined length polymorphisms are particularly useful in screening processes.

In another embodiment, the invention provides an improved method that uses PCR amplification of a genomic HLA DNA sequence of one HLA locus. Following amplification, the amplified DNA sequence is combined with at least one endonuclease to produce a digest. The endonuclease cleaves the amplified DNA sequence to yield a set of fragments having distinctive fragment lengths. Usually the amplified sequence is divided, and two or more endonuclease digests are produced. The digests can

be used, either separately or combined, to produce RFLP patterns that can distinguish between individuals.

Additional digests can be prepared to provide enhanced specificity to distinguish between even closely related individuals with the same HLA type.

In a preferred embodiment, the presence of a particular allele can be verified by performing a two step amplification procedure in which an amplified sequence produced by a first primer pair is amplified by a second primer pair which binds to and defines a sequence within the first amplified sequence. The first primer pair can be specific for one or more alleles of the HLA locus. The second primer pair is preferably specific for one allele of the HLA locus, rather than a plurality of alleles. The presence of an amplified sequence indicates the presence of the allele, which is confirmed by production of characteristic RFLP patterns.

To analyze RFLP patterns, fragments in the digest are separated by size and then visualized. In the case of typing for a particular HLA locus, the analysis is directed to detecting the two DNA allele sequences that uniquely characterize that locus in each individual. Usually this is performed by comparing the sample digest RFLP patterns to a pattern produced by a control sample of known HLA allele type. However, when the method is used for paternity testing or forensics, the analysis need not involve identifying a particular locus or loci but can be done by comparing single or multiple RFLP patterns of one individual with that of another individual using the same restriction endonuclease and primers to determine similarities and differences between the patterns.

The number of digests that need to be prepared for any particular analysis will depend on the desired information and the particular sample to be analyzed. For example, one digest may be sufficient to determine

that an individual cannot be the person whose blood was found at a crime scene. In general, the use of two to three digests for each of two to three HLA loci will be sufficient for matching applications (forensics, 5 paternity). For complete HLA haplotyping; e.g., for transplantation, additional loci may need to be analyzed.

As described previously, combinations of primer pairs can be used in the amplification method to amplify 10 a particular HLA DNA locus irrespective of the allele present in the sample. In a preferred embodiment, samples of HLA DNA are divided into aliquots containing similar amounts of DNA per aliquot and are amplified 15 with primer pairs (or combinations of primer pairs) to produce amplified DNA sequences for additional HLA loci. Each amplification mixture contains only primer pairs 20 for one HLA locus. The amplified sequences are preferably processed concurrently, so that a number of digest RFLP fragment patterns can be produced from one sample. In this way, the HLA type for a number of alleles can be determined simultaneously.

Alternatively, preparation of a number of RFLP fragment patterns provides additional comparisons of 25 patterns to distinguish samples for forensic and paternity analyses where analysis of one locus frequently fails to provide sufficient information for the determination when the sample DNA has the same allele as the DNA to which it is compared.

The use of HLA types in paternity tests or 30 transplantation testing and in disease diagnosis and prognosis is described in Basic & Clinical Immunology, 3rd Ed (1980) Lange Medical Publications, pp 187-190, which is incorporated herein by reference in its 35 entirety. HLA determinations fall into two general categories. The first involves matching of DNA from an individual and a sample. This category involves

forensic determinations and paternity testing. For category 1 analysis, the particular HLA type is not as important as whether the DNA from the individuals is related. The second category is in tissue typing such as for use in transplantation. In this case, rejection of the donated blood or tissue will depend on whether the recipient and the donor express the same or different antigens. This is in contrast to first category analyses where differences in the HLA DNA in either the introns or exons is determinative.

For forensic applications, analysis of the sample DNA of the suspected perpetrator of the crime and DNA found at the crime scene are analyzed concurrently and compared to determine whether the DNA is from the same individual. The determination preferably includes analysis of at least three digests of amplified DNA of the DQA1 locus and preferably also of the A locus. More preferably, the determination also includes analysis of at least three digests of amplified DNA of an additional locus, e.g. the DPB locus. In this way, the probability that differences between the DNA samples can be discriminated is sufficient.

For paternity testing, the analysis involves comparison of DNA of the child, the mother and the putative father to determine the probability that the child inherited the obligate haplotype DNA from the putative father. That is, any DNA sequence in the child that is not present in the mother's DNA must be consistent with being provided by the putative father. Analysis of two to three digests for the DQA1 and preferably also for the A locus is usually sufficient. More preferably, the determination also includes analysis of digests of an additional locus, e.g. the DPB locus.

For tissue typing determinations for transplantation matching, analysis of three loci (HLA A,

B, and DR) is often sufficient. Preferably, the final analysis involves comparison of additional loci including DQ and DP.

5       Production of RFLP fragment patterns

The following table of exemplary fragment pattern lengths demonstrates distinctive patterns. For example, as shown in the table, Bsrl cleaves A2, A3 and A9 allele amplified sequences defined by primers SGD005.IIVS1.LNP and SGD009.AIVS3.R2NP into sets of fragments with the following numbers of nucleotides (740, 691), (809, 335, 283) and (619, 462, 256, 93), respectively. The fragment patterns clearly indicate which of the three A alleles is present. The following table illustrates a number of exemplary endonucleases that produce distinctive RFLP fragment patterns for exemplary A allele sequences.

Table 2 illustrates the set of RFLP fragments produced by use of the designated endonucleases for analysis of three A locus alleles. For each endonuclease, the number of nucleotides of each of the fragments in a set produced by the endonuclease is listed. The first portion of the table illustrates RFLP fragment lengths using the primers designated SGD009.AIVS3.R2NP and SGD005.IIVS1.LNP which produce the longer of the two exemplary sequences. The second portion of the table illustrates RFLP fragment lengths using the primers designated SGD006.AIVS3.R1NP and SGD005.IIVS1.LNP which produce the shorter of the sequences. The third portion of the table illustrates the lengths of fragments of a DQA1 locus-specific amplified sequence defined by the primers designated SGD001.DQA1.LNP and SGD003.DQA1.RNP.

As shown in the Table, each of the endonucleases produces a characteristic RFLP fragment pattern which can readily distinguish which of the three A alleles is

present in a sample.

169.0018

169.0018

TABLE 5

## RFLP FRAGMENT PATTERNS

5

## A - Long

	BsrI	A2	740	691				
10		A3	809					
		A9		619	462	335	283	
	Cfr101	A2	1055		399	245		
15		A3		473	399	247		
		A9	786		399			
	DraII	A2	698			251		138
20		A3		369	315	251	247	
		A9	596	427		251		80
25	FokI	A2	728	248		151		
		A3		515	225	213	151	
		A9	1004			151		
30	GsuI	A2	868	547			36	
		A3	904		523			
		A9		638		419	373	
35	HphI	A2	1040		239			72
		A3		419	375			
		A9	643	419	373			
40	MboII	A2	1011	165	143	132		
		A3		893	194			
		A9	1349		143		115	51
45	PpumI	A2	698		295	251		138
		A3		369	364	251	242	
		A9	676	503		251		
	PssI	A2	695		295	251		138
		A3		366	315	251	242	
		A9	596	427		251		

**A - Short**

5	BsrI	A2	691			254		
		A3		345	335	283		
		A9	619				256	93
10	Cfr101	A2						
		A3						
		A9						
15	DraII	A2		295	251	210	138	
		A3	315		251	210		
		A9	427		251	210		
	FokI	A2	293	248			151	143
		A3			225	213	151	143
20		A9	539				151	146
	GsuI	A2	868			61	36	
		A3	904			59		
		A9		414	373	178		
25	HphI	A2	554		339			
		A3		411	375		177	
		A9		414	373		178	
30	MboII	A2						
		A3						
		A9						
35	PpumI	A2		295	257		212	69
		A3	364			251	210	72
		A9	503			251	211	66
	PssI	A2		295	251	219		
		A3	315		251		207	72
40		A9	427		251		208	72

45

DOA 1

Screening Analysis for Genetic Disease

Carriers of genetic diseases and those affected by the disease can be identified by use of the present method. Depending on the disease, the screening analysis can be used to detect the presence of one or more alleles associated with the disease or the presence of haplotypes associated with the disease. Furthermore, by analyzing haplotypes, the method can detect genetic diseases that are not associated with coding region variations but are found in regulatory or other untranslated regions of the genetic locus. The screening method is exemplified below by analysis of cystic fibrosis (CF).

Cystic fibrosis is an autosomal recessive disease, requiring the presence of a mutant gene on each chromosome. CF is the most common genetic disease in Caucasians, occurring once in 2,000 live births. It is estimated that one in forty Caucasians are carriers for the disease.

Recently a specific deletion of three adjacent basepairs in the open reading frame of the putative CF gene leading to the loss of a phenylalanine residue at position 508 of the predicted 1480 amino acid polypeptide was reported [Kerem et al, *Science* **245**:1073-1080 (1989)]. Based on haplotype analysis, the deletion may account for most CF mutations in Northern European populations (about 68%). A second mutation is reportedly prevalent in some Southern European populations. Additional data indicate that several other mutations may cause the disease.

Studies of haplotypes of parents of CF patients (who necessarily have one normal and one disease-associated haplotype) indicated that there are at least 178 haplotypes associated with the CF locus. Of those haplotypes, 90 are associated only with the disease; 78 are found only in normals; and 10 are associated with

both the disease and with normals (Kerem et al, supra).  
The disease apparently is caused by several different mutations, some in very low frequency in the population.  
As demonstrated by the haplotype information, there are more haplotypes associated with the locus than there are mutant alleles responsible for the disease.

A genetic screening program (based on amplification of exon regions and analysis of the resultant amplified DNA sequence with probes specific for each of the mutations or with enzymes producing RFLP patterns characteristic of each mutation) may take years to develop. Such tests would depend on detection and characterization of each of the mutations, or at least of mutations causing about 90 to 95% or more of the cases of the disease. The alternative is to detect only 70 to 80% of the CF-associated genes. That alternative is generally considered unacceptable and is the cause of much concern in the scientific community.

The present method directly determines haplotypes associated with the locus and can detect haplotypes among the 178 currently recognized haplotypes associated with the disease locus. Additional haplotypes associated with the disease are readily determined through the rapid analysis of DNA of numerous CF patients by the methods of this invention. Furthermore, any mutations which may be associated with noncoding regulatory regions can also be detected by the method and will be identified by the screening process.

Rather than attempting to determine and then detect each defect in a coding region that causes the disease, the present method amplifies intron sequences associated with the locus to determine allelic and sub-allelic patterns. In contrast to use of mutation-specific probes where only known sequence defects can be detected, new PDLP and RFLP patterns produced by intron

sequences indicate the presence of a previously unrecognized haplotype.

The same analysis can be performed for phenylalanine hydroxylase locus mutations that cause phenylketonuria and for beta-globin mutations that cause beta-thalassemia and sickle cell disease and for other loci known to be associated with a genetic disease. Furthermore, neither the mutation site nor the location for a disease gene is required to determine haplotypes associated with the disease. Amplified intron sequences in the regions of closely flanking RFLP markers, such as are known for Huntington's disease and many other inherited diseases, can provide sufficient information to screen for haplotypes associated with the disease.

Muscular dystrophy (MD) is a sex-linked disease. The disease-associated gene comprises a 2.3 million basepair sequence that encodes 3,685 amino acid protein, dystrophin. A map of mutations for 128 of 34 patients with Becker's muscular dystrophy and 160 patients with Duchenne muscular dystrophy identified 115 deletions and 13 duplications in the coding region sequence [Den Dunnen et al, *Am. J. Hum. Genet.* **45**:835-847 (1989)]. Although the disease is associated with a large number of mutations that vary widely, the mutations have a non-random distribution in the sequence and are localized to two major mutation hot spots, Den Dunnen et al, *supra*. Further, a recombination hot spot within the gene sequence has been identified [Grimm et al, *Am. J. Hum. Genet.* **45**:368-372 (1989)].

For analysis of MD, haplotypes on each side of the recombination hot spot are preferably determined. Primer pairs defining amplified DNA sequences are preferably located near, within about 1 to 10 Kbp of the hot spot on either side of the hot spot. In addition, due to the large size of the gene, primer pairs defining amplified DNA sequences are preferably located near each

end of the gene sequence and most preferably also in an intermediate location on each side of the hot spot. In this way, haplotypes associated with the disease can be identified.

5 Other diseases, particularly malignancies, have been shown to be the result of an inherited recessive gene together with a somatic mutation of the normal gene. One malignancy that is due to such "loss of heterogeneity" is retinoblastoma, a childhood cancer.  
10 The loss of the normal gene through mutation has been demonstrated by detection of the presence of one mutation in all somatic cells (indicating germ cell origin) and detection of a second mutation in some somatic cells [Scheffer et al, *Am. J. Hum. Genet.*  
15 45:252-260 (1989)]. The disease can be detected by amplifying somatic cell, genomic DNA sequences that encompass sufficient intron sequence nucleotides. The amplified DNA sequences preferably encompass intron sequences locate near one or more of the markers described by Scheffer et al, *supra*. Preferably, an amplified DNA sequence located near an intragenic marker and an amplified DNA sequence located near a flanking marker are used.  
20

25 An exemplary analysis for CF is described in detail in the examples. Analysis of genetic loci for other monogenic and multigenic genetic diseases can be performed in a similar manner.

As the foregoing description indicates, the present method of analysis of intron sequences is generally applicable to detection of any type of genetic trait. Other monogenic and multigenic traits can be readily analyzed by the methods of the present invention. Furthermore, the analysis methods of the present method are applicable to all eukaryotic cells, and are preferably used on those of plants and animals. Examples of analysis of BoLA (bovine MHC determinants)

further demonstrates the general applicability of the methods of this invention.

This invention is further illustrated by the following specific but non-limiting examples. Procedures that are constructively reduced to practice are described in the present tense, and procedures that have been carried out in the laboratory are set forth in the past tense.

10

#### EXAMPLE 1

##### Forensic Testing

15

DNA extracted from peripheral blood of the suspected perpetrator of a crime and DNA from blood found at the crime scene are analyzed to determine whether the two samples of DNA are from the same individual or from different individuals.

20

The extracted DNA from each sample is used to form two replicate aliquots per sample, each aliquot having 1 µg of sample DNA. Each replicate is combined in a total volume of 100 µl with a primer pair (1 µg of each primer), dNTPs (2.5 mM each) and 2.5 units of Taq polymerase in amplification buffer (50 mM KCl; 10 mM Tris-HCl, pH 8.0; 2.5 mM MgCl<sub>2</sub>; 100 µg/ml gelatin) to form four amplification reaction mixtures. The first primer pair contains the primers designated SGD005.IIIVS1.LNP and SGD009.AIVS3.R2NP (A locus-specific). The second primer pair contains the primers designated SGD001.DQA1.LNP and SGD003.DQA1.RNP (DQA locus-specific). Each primer is synthesized using an Applied Biosystems model 308A DNA synthesizer. The amplification reaction mixtures are designated SA (suspect's DNA, A locus-specific primers), SD (suspect's DNA, DQA1 locus-specific primers), CA (crime scene DNA, A locus-specific primers) and CD (crime scene DNA, DQA1 locus-specific primers).

30

35

Each amplification reaction mixture is heated to 94°C for 30 seconds. The primers are annealed to the sample DNA by cooling the reaction mixtures to 65°C for each of the A locus-specific amplification mixtures and to 55°C for each of the DQA1 locus-specific amplification mixtures and maintaining the respective temperatures for one minute. The primer extension step is performed by heating each of the amplification mixtures to 72°C for one minute. The denaturation, annealing and extension cycle is repeated 30 times for each amplification mixture.

Each amplification mixture is aliquoted to prepare three restriction endonuclease digestion mixtures per amplification mixture. The A locus reaction mixtures are combined with the endonucleases BsrlI, Cfr101 and DraII. The DQA1 reaction mixtures are combined with AluI, CviJI and DdeI.

To produce each digestion mixture, each of three replicate aliquots of 10 µl of each amplification mixture is combined with 5 units of the respective enzyme for 60 minutes at 37°C under conditions recommended by the manufacturer of each endonuclease.

Following digestion, the three digestion mixtures for each of the samples (SA, SD, CA and CD) are pooled and electrophoresed on a 6.5% polyacrylamide gel for 45 minutes at 100 volts. Following electrophoresis, the gel is stained with ethidium bromide.

The samples contain fragments of the following lengths:

30

SA: 786, 619, 596, 462, 427, 399, 256, 251, 93, 80  
CA: 809, 786, 619, 596, 473, 462, 427, 399, 369, 335,  
315, 283, 256, 251, 247, 93, 80

5 SD: 388, 338, 332, 277, 219, 194, 122, 102, 89, 79,  
64, 55  
CD: 587, 449, 388, 338, 335, 332, 277, 271, 219, 194,  
187, 122, 102, 99, 89, 88, 79, 65, 64, 55

10 The analysis demonstrates that the blood from the crime scene and from the suspected perpetrator are not from the same individual. The blood from the crime scene and from the suspected perpetrator are, respectively, A3, A9, DQA1 0501, DQA1 0301 and A9, A9,  
15 DQA1 0501, DQA1 0501.

#### EXAMPLE 2

##### Paternity Testing

20 Chorionic villus tissue was obtained by trans-cervical biopsy from a 7-week old conceptus (fetus). Blood samples were obtained by venepuncture from the mother (M), and from the alleged father (AF). DNA was extracted from the chorionic villus biopsy, and from the blood samples. DNA was extracted from the sample from M by use of nonionic detergent (Tween 20) and proteinase K. DNA was extracted from the sample from F by hypotonic lysis. More specifically, 100  $\mu$ l of blood was diluted to 1.5 ml in PBS and centrifuged to remove buffy coat. Following two hypotonic lysis treatments  
25 involving resuspension of buffy coat cells in water, the pellets were washed until redness disappeared. Colorless pellets were resuspended in water and boiled for 20 minutes. Five 10 mm chorionic villus fronds were received. One frond was immersed in 200  $\mu$ l water. NaOH  
30 was added to 0.05 M. The sample was boiled for 20  
35

minutes and then neutralized with HCl. No further purification was performed for any of the samples.

The extracted DNA was submitted to PCR for amplification of sequences associated with the HLA loci, DQA1 and DPB1. The primers used were: (1) as a 5' primer for the DQA1 locus, the primer designated SGD001.DQA1.LNP (DQA 5'IVS1) (corresponding to nt 23-39 of the DQA1 0301 allele sequence) and as the 3' primer for the DQA1 locus, the primer designated SGD003.DQA1.RNP (DQA 3'IVS2 corresponding to nt 789-806 of the DQA1 0301 sequence; (2) as the DPB primers, the primers designated 5'IVS1 nt 7604-7624 and 3'IVS2 7910-7929. The amplification reaction mixtures were: 150 ng of each primer; 25  $\mu$ l of test DNA; 10 mM Tris HCl, pH 8.3; 50 mM KCl; 1.5 mM MgCl<sub>2</sub>; 0.01% (w/v) gelatin; 200  $\mu$ M dNTPs; water to 100  $\mu$ l and 2.5 U Taq polymerase.

The amplification was performed by heating the amplification reaction mixture to 94°C for 10 minutes prior to addition of Taq polymerase. For DQA1, the amplification was performed at 94°C for 30 seconds, then 55°C for 30 seconds, then 72°C for 1 minute for 30 cycles, finishing with 72°C for 10 minutes. For DPB, the amplification was performed at 96°C for 30 seconds, then 65°C for 30 seconds, finishing with 65°C for 10 minutes.

Amplification was shown to be technically satisfactory by test gel electrophoresis which demonstrated the presence of double stranded DNA of the anticipated size in the amplification reaction mixture. The test gel was 2% agarose in TBE (tris borate EDTA) buffer, loaded with 15  $\mu$ l of the amplification reaction mixture per lane and electrophoresed at 200 v for about 2 hours until the tracker dye migrated between 6 to 7 cm into the 10 cm gel.

The amplified DQA1 and DPB1 sequences were subjected to restriction endonuclease digestion using

DdeI and MboII (8 and 12 units, respectively at 37°C for 3 hours) for DQA1, and RsaI and FokI (8 and 11 units, respectively at 37°C overnight) for DPB1 in 0.5 to 2.0 µl of enzyme buffers recommended by the supplier, Pharmacia together with 16-18 µl of the amplified product. The digested DNA was fragment size-length separated on gel electrophoresis (3% Nusieve). The RFLP patterns were examined under ultraviolet light after staining the gel with ethidium bromide.

Fragment pattern analysis is performed by allele assignment of the non-maternal alleles using expected fragment sizes based on the sequences of known endonuclease restriction sites. The fragment pattern analysis revealed the obligate paternal DQA1 allele to be DQA1 0102 and DPB to be DPB1. The fragment patterns were consistent with AF being the biological father.

To calculate the probability of true paternity, HLA types were assigned. Maternal and AF DQA1 types were consistent with those predicted from the HLA Class II gene types determined by serological testing using lymphocytotoxic antisera.

Considering alleles of the two HLA loci as being in linkage equilibrium, the combined probability of non-paternity was given by:

$$0.042 \times 0.314 = 0.013$$

i.e. the probability of paternity is  $(1 - 0.013)$  or 98.7%.

The relative chance of paternity is thus 74:75, i.e. the chance that the AF is not the biological father is approximately 1 in 75. The parties to the dispute chose to regard these results as confirming the paternity of the fetus by the alleged father.

### EXAMPLE 3

#### Analysis of the HLA DQA1 Locus

The three haplotypes of the HLA DQA1 0102 locus were analyzed as described below. Those haplotypes are 5 DQA1 0102 DR15 Dw2; DQA1 0102 DR16 Dw21; and DQA1 0102 DR13 Dw19. The distinction between the haplotypes is particularly difficult because there is a one basepair difference between the 0102 alleles and the 0101 and 10 0103 alleles, which difference is not unique in DQA1 allele sequences.

The procedure used for the amplification is the same as that described in Example 1, except that the amplification used thirty cycles of 94°C for 30 seconds, 15 60°C for 30 seconds, and 72°C for 60 seconds. The sequences of the primers were:

SGD 001 -- 5' TTCTGAGCCAGTCCTGAGA 3'; and  
SGD 003 -- 5' GATCTGGGGACCTCTTGG 3'.

These primers hybridize to sequences about 500 bp upstream from the 5' end of the second exon and 50 bp downstream from the second exon and produce amplified DNA sequences in the 700 to 800 bp range. 20

Following amplification, the amplified DNA sequences were electrophoresed on a 4% polyacrylamide gel to determine the PDLA type. In this case, amplified DNA sequences for 0102 comigrate with (are the same length as) 0101 alleles and subsequent enzyme digestion 25 is necessary to distinguish them.

The amplified DNA sequences were digested using the restriction enzyme AluI (Bethesda Research Laboratories) which cleaves DNA at the sequence AGCT. 30 The digestion was performed by mixing 5 units (1 µl) of enzyme with 10 µl of the amplified DNA sequence (between about 0.5 and 1 µg of DNA) in the enzyme buffer provided by the manufacturer according to the manufacturer's directions to form a digest. The digest was then 35

incubated for 2 hours at 37°C for complete enzymatic digestion.

The products of the digestion reaction are mixed with approximately 0.1 µg of "ladder" nucleotide sequences (nucleotide control sequences beginning at 5 123 bp in length and increasing in length by 123 bp to a final size of about 5,000 bp; available commercially from Bethesda Research Laboratories, Bethesda MD) and were electrophoresed using a 4% horizontal ultra-thin 10 polyacrylamide gel (E-C Apparatus, Clearwater FLA). The bands in the gel were visualized (stained) using 15 silver stain technique [Allen et al, *BioTechniques* 7:736-744 (1989)].

Three distinctive fragment patterns which 15 correspond to the three haplotypes were produced using AluI. The patterns (in base pair sized fragments) were:

1. DR15 DQ6 Dw2: 120, 350, 370, 480
2. DR13 DQ6 Dw19: 120, 330, 350, 480
3. DR16 DQ6 Dw21: 120, 330, 350

The procedure was repeated using a 6.5% vertical 20 polyacrylamide gel and ethidium bromide stain and provided the same results. However, the fragment 25 patterns were more readily distinguishable using the ultrathin gels and silver stain.

This exemplifies analysis according to the method of this invention. Using the same procedure, 20 of the other 32 DR/DQ haplotypes for DQA1 were identified using the same primer pair and two additional enzymes (DdeI 30 and MboII). PDLP groups and fragment patterns for each of the DQA1 haplotypes with the three endonucleases are illustrated in Table 6.

AluI													
PDLP	DR	Dw	480	405	390	360	340	310	270	240	120	100	
			410	400	370	350	330	300	270	240	110		
0101	1	1								+	+	+	
	1	20								+	+	+	
	14	9								+	+	+	
0102	15	2		+		+	+					+	
	16	21		+		+	+					+	
	13	19		+		+	+					+	
0103	13	18, 24											
	13	18, 25											
	8	8.3											
	11	DB2											
	15	12											
0201	7	DB1									+		
	7	17									+		
	7	11									+		
0301	4	4(7)											
	4	13.2(7)											
	4	4(8)											
	4	10(8)											
	4	13.1(8)											
	4	14(8)											
	4	KT2											
	4	15											
	9	23					+						
0401	8	8.1		(+)							+	+	+
	8	8.2		(+)							+	+	+
	3	RSH											
0501	3	3, 24		(+)							+		
	3	3, 25		(+)							+		
	11	5		(+)							+		
	11	5(9104)		(+)							+		
	14	16		(+)							+		
	12	DB6		(+)							+		
0601	16	22		(+)							+		
	8	8.3		(+)							+		
PDLP	DR	Dw	480	405	390	360	340	330	300	270	240	110	100

			DdeI															
	PDLP	DR	Dw	650	520	450	420	410	400	390	300	200	190	150	90	80	60	50
5	0101	1	1						+					+	+	+		
		1	20						+					+	+	+		
		14	9						+					+	+	+		
10	0102	15	2					+	+					+	+	+		
		16	21						+					+	+	+		
		13	19						+					+	+	+		
15	0103	13	18,24															
		13	18,25															
		8	8.3						+									
20	0201	11	DB2															
		15	12															
		7	DB1											+	+	+	+	
25	0301	7	17											+	+	+	+	
		7	11											+	+	+	+	
		4	4(7)															
30	0301	4	13.2(7)															
		4	4(8)						+									
		4	10(8)						+									
		4	13.1(8)															
		4	14(8)															
		4	KT2															
		4	15															
		9	23						+									
		8	8.1											+	+	+	+	
35	0401	8	8.2											+	+	+	+	
		3	RSH											+	+	+	+	
		3	3,24											+	+	+	+	
40	0501	3	3,25											+	+	+	+	
		11	5											+	+	+	+	
		11	5(9104)											+	+	+	+	
		14	16											+	+	+	+	
		12	DB6											+	+	+	+	
		16	22											+	+	+	+	
		8	8.3											+	+	+	+	
45	PDLP	DR	Dw	650	520	450	420	410	400	390	300	200	190	150	90	80	60	50

all other genes have been mapped to the same chromosome. In some cases, several genes are located on the same chromosome.

5  
10  
15  
20  
25  
30  
35  
40  
45  
50

			MboII															
PDLP	DR	Dw	390	385	380	370	365	360	350	340	335	330	305	300	250	190	170	140
			390	385	380	370	365	360	350	340	335	330	305	300	250	190	170	140
0101	1	1							+						+			
	1	20							+						+			
	14	9							+						+			
0102	15	2							+						+			
	16	21							+						+			
	13	19							+						+			
0103	13	18,24																
	13	18,25																
	8	8.3																
	11	DB2																
	15	12																
0201	7	DB1								+								
	7	17							+						+			
	7	11							+						+			
0301	4	4(7)																
	4	13.2(7)																
	4	4(8)													+			
	4	10(8)													+			
	4	13.1(8)													+			
	4	14(8)													+			
	4	KT2													+			
	4	15													+			
	9	23													+			
0401	8	8.1								+								
	8	8.2							+									
	3	RSH																
0501	3	3,24							+									
	3	3,25							+									
	11	5							+									
	11	5(9104)							+									
	14	16							+									
	12	DB6							+									
	16	22							+									
0601	8	8.3												+				
PDLP	DR	Dw		385	380	370	365	360	350	340	335	330	305	300	250	190	170	140
				390	380	365	350	335	330	305	300	250	190	170	140			

This example illustrates the ability of the method of this invention to distinguish the alleles and haplotypes of a genetic locus. Specifically, the example shows that PDLP analysis stratifies five of the eight alleles. These three restriction endonuclease digests distinguish each of the eight alleles and many of the 35 known haplotypes of the locus. The use of additional endonuclease digests for this amplified DNA sequence can be expected to distinguish all of the known haplotypes and to potentially identify other previously unrecognized haplotypes. Alternatively, use of the same or other endonuclease digests for another amplified DNA sequence in this locus can be expected to distinguish the haplotypes.

In addition, analysis of amplified DNA sequences at the DRA locus in the telomeric direction and DQB in the centromeric direction, preferably together with analysis of a central locus, can readily distinguish all of the haplotypes for the region.

The same methods are readily applied to other loci.

#### EXAMPLE 4

##### Analysis of the HLA DQA1 Locus

The DNA of an individual is analyzed to determine which of the three haplotypes of the HLA DQA1 0102 locus are present. Genomic DNA is amplified as described in Example 3. Each of the amplified DNA sequences is sequenced to identify the haplotypes of the individual. The individual is shown to have the haplotypes DR15 DQ6 Dw2; DR13 DQ6 Dw19.

The procedure is repeated as described in Example 3 through the production of the AluI digest. Each of the digest fragments is sequenced. The individual is shown to have the haplotypes DR15 DQ6 Dw2; DR13 DQ6 Dw19.

EXAMPLE 5

DQA1 Allele-Specific Amplification

Primers were synthesized that specifically bind the 0102 and 0301 alleles of the DQA1 locus. The 5' 5 primer was the SGD 001 primer used in Example 3. The sequences of the 3' primers are listed below.

0102 5' TTGCTGAACTCAGGCCACC 3'

0301 5' TGCAGAACAGAGGCCACTG 3'

The amplification was performed as described in Example 10 3 using 30 cycles of a standard (94°C, 60°C, 72°C) PCR reaction. The template DNAs for each of the 0101, 0301 and 0501 alleles were amplified separately. As determined by gel electrophoresis, the 0102-allele-specific 15 primer amplified only template 0102 DNA and the 0301-allele-specific primer amplified only template 0301 DNA. Thus, each of the primers was allele-specific.

EXAMPLE 6

Detection of Cystic Fibrosis

The procedure used for the amplification described 20 in Example 3 is repeated. The sequences of the primers are illustrated below. The first two primers are upstream primers, and the third is a downstream primer. 25 The primers amplify a DNA sequence that encompasses all of intervening sequence 1

5' CAG AGG TCG CCT CTG GA 3';

5' AAG GCC AGC GTT GTC TCC A 3'; and

3' CCT CAA AAT TGG TCT GGT 5'.

30 These primers hybridize to the complement of sequences located from nt 136-152 and nt 154-172, and to nt 187-207. [The nucleotide numbers are found in Riordan et al, *Science* 245:1066-1072 (1989).]

35 Following amplification, the amplified DNA sequences are electrophoresed on a 4% polyacrylamide gel to determine the PDLP type. The amplified DNA sequences

are separately digested using each of the restriction enzymes AluI, MnII and RsaI (Bethesda Research Laboratories). The digestion is performed as described in Example 3. The products of the digestion reaction are electrophoresed and visualized using a 4% horizontal ultra-thin polyacrylamide gel and silver stain as described in Example 3.

Distinctive fragment patterns which correspond to disease-associated and normal haplotypes are produced.

### EXAMPLE 7

## Analysis of Bovine HLA Class I

Bovine HLA Class I alleles and haplotypes are analyzed in the same manner as described in Example 3. The primers are listed below.

The primers are listed below.

Bovine Primers (Class I HLA homolog)	$T_m$
5' primer: 5' TCC TGG TCC TGA CCG AGA 3'	(62°)
3' primer: 1) 3' A TGT GCC TTT GGA GGG TCT 5'	(62°)
(for ~600 bp product)	
2) 3' GCC AAC AT GAT CCG CAT 5'	(62°)
(for ~900 bp product)	

For the approximately 900 bp sequence PDLP analysis is sufficient to distinguish alleles 1 and 3 (893 and 911 bp, respectively). Digests are prepared as described in Example 3 using AluI and DdeI. The following patterns are produced for the 900 bp sequence.

Allele 1: AluT digest: 712, 181

Allele 3, AluT digest: 430, 300, 181

Allele 1: Pdet digest: 445 201 182 28

Allele 3. PdeI digest: 406, 185, 182, 28, 16

The 600 bp sequence also produces distinguishable fragment patterns for those alleles. However, those

patterns are not as dramatically different as the patterns produced by the 600 bp sequence digests.

#### EXAMPLE 8

5

##### Preparation of Primers

Each of the following primers is synthesized using an Applied Biosystems model 308A DNA synthesizer.

###### HLA locus primers

###### A locus-specific primers

10 SGD009.AIVS3.R2NP CATGTGGCCATCTTGAGAATGGA  
SGD006.AIVS3.R1NP GCCCGGGAGATCTACAGGCGATCA  
A2.1 CGCCTCCCTGATCGCCTGTAG  
A2.2 CCAGAGAGTGAUTCTGAGG  
A2.3 CACAATTAAGGGAT

15

###### B locus-specific primers

SGD007.BIVS3.R1NP TCCCCGGCGACCTATAAGGAGATGG  
SGD010.BIVS3.R2NP CTAGGACCACCCATGTGACCAGC  
B2.1 ATCTCCTCAGACGCCGAGATGCGTCAC  
B2.2 CTCCTGCTGCTCTGGGGGGCAG  
B2.3 ACTTTACCTCCACTCAGATCAGGAG  
B2.4 CGTCCAGGCTGGTGTCTGGTTCTGTGCCCT  
B2.5 CTGGTCACATGGGTGGTCCTAGG  
B2.6 CGCCTGAATTCTGACTCTCCCAT

25

###### C locus-specific primers

SGD008.CIVS3.R1NP ATCCCAGGAGATCTACAGGAGATG  
SGD011.CIVS3.R2NP AACAGCGCCCATGTGACCATCCT  
C2.1 CTGGGGAGGCGCCGCGTTGAGGATTCT  
C2.2 CGTCTCCGCAGTCCCAGTTCTAAAGTTCCCAGT  
C2.3 ATCCTCGTGCTCTCGGGA  
C2.4 TGTGGTCAGGCTGCTGAC  
C2.5 AAGGTTGATTCCAGCTT  
C2.6 CCCCTCCCCACCCAGGTGTTCCGTCCATTCTTCAGGA  
C2.7 CACATGGGCCTGTTGGAGTGTG

Class I loci-specific primers

SGD005.IIVS1.LNP GTGAGTGCAGGGTCGGGAGGGA

1.1 CACCCACCGGGACTCAGA

1.2 TGGCCCTGACCCAGACCTGGGC

5 1.3 GAGGGTCGGCGGGTCTCAGC

1.4 CTCTCAGGCCTTGTTC

1.5 CAGAAGTCGCTGTTCC

DQA1 locus-specific primers

10 SGD001.DQA1.LNP TTCTGAGCCAGTCCTGAGA

DQA3 E1a TTGCCCTGACCACCGTGATG

DQA3 E1b CTTCCCTGCTTGTCACTTCA

DQA3 E1c CCATGAATTGATGGAGA

DQA3 E1d ACCGCTGCTACCAATGGTA

15 SGD003.DQA1.RNP CCAAGAGGTCCCCAGATC

DRA locus-specific primers

DRA E1 TCATCATAGCTGTGCTGATG

DRA 5'E2 AGAACATGTGATCATCCAGGC

20 DRA 3'E2 CCAACTATACTCCGATCACCAAT

DRB locus-specific primers

DRB E1 TGACAGTGACACTGATGGTGCTG

DRB 5'E2 GGGGACACCCGACCACGTTTC

25 DRB 3'E2 TGCAGACACAACACGGGTTG

DOB1 locus-specific primers

DQB E1 TGGCTGAGGGCAGAGACTCTCCC

DQB 5'E2 TGCTACTTCACCAACGGGAC

30 DQB 3'E2 GGTGTGCACACACAAC

DQB 5'IVS1a AGGTATTTACCCAGGGACCAAGAGAT

DQB 5'IVS1b ATGTAAAATCAGCCCGACTGCCTCTTC

DQB 3'IVS2 GCCTCGTGCCTTATGCGTTGCCTCCT

35 DPB1 locus-specific primers

DPB E1 TGAGGTTAATAACTGGAGAA

DPB 5' IVS1 GAGAGTGGCGCCTCCGCTCAT  
DPB 3' IVS2 GAGTGAGGGCTTGGGCCGG